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(54) Title: PROCESSES OF MAKING NORTH AMERICAN GINSENG FRACTIONS, PRODUCTS CONTAINING THEM, AND USE AS IMMUNOMODULATORS (57) Abstract The invention is directed to chemical processes of preparing fractions from North American ginseng (<i>Panax quinquefolium</i>) and pharmaceutical compositions containing these fractions. The products of the present invention may be used to stimulate the production of cytokines and/or antibodies, or as therapeutics targeted at conditions characterized by low immunity, such as the common cold, influenza, chronic fatigue syndrome, AIDS and cancer.		

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**PROCESSES OF MAKING NORTH AMERICAN GINSENG
FRACTIONS,
PRODUCTS CONTAINING THEM, AND USE AS
IMMUNOMODULATORS**

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Field of the Invention

This invention relates to chemical processes of making fractions from North American ginseng (*Panax quinquefolium*) and compositions containing these fractions. The products of the present invention may be used to stimulate the production of antibodies, or as
10 therapeutics targeted at conditions characterized by low immunity, such as the common cold, influenza, chronic fatigue syndrome, AIDS, cancer, etc. The products of the present invention may also be used as a supplement for cancer patients undergoing chemotherapy or radiation therapy, which is known to cause serious suppression of the immune
15 system.

Background of the Invention

For hundreds of years, the use of certain non-toxic agents such as herbal compounds has been widely accepted for a variety of physiological conditions, especially in the Orient. *Panax ginseng* C.A.
20 Meyer is the best known traditional Chinese medicine. The important pharmacological activities of ginseng extracts, alone or in combination with other drugs, include alleviation of renal impairment, inhibition of carcinogenesis and prevention of stress. There are also a number of reports on the influence of ginseng on the immunological
25 responsiveness of the individual. Some immunomodulatory properties that have been reported include enhancement of host resistance against infection, anti-inflammatory effect, inhibition of tumor growth, as well as modulation of some basic immune function at the cellular level. American ginseng, *Panax quinquefolium*, is another specie of ginseng

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which has gained popularity as a health supplement having many beneficial health effects. Several groups of scientists have attempted to isolate and elucidate the structure of the polysaccharides present in ginseng. Some of the polysaccharides have been demonstrated to be
5 active in modulating the immune system.

A series of studies on the isolation, characterization, and biological evaluation of ginseng polysaccharides was carried out by Tomoda's group in Kyoritsu College of Pharmacy, Japan. In one set of studies, ginseng polysaccharides were fractionated based on their
10 acidity. Two acidic polysaccharides having immunological activities have been isolated from root of Korean ginseng (*Panax ginseng*)^[1,2]. The sliced roots were extracted with hot water. The extract was treated with cetyltrimethylammonium bromide (CTAB) in the presence of sodium sulfate. The precipitate was separated, dialyzed, and applied to a
15 Sephadex G-25 column, DEAE-Sephacel (Pharmacia) column to give two pure polysaccharides, which were designated as ginsenan PA and ginsenan PB. Gel chromatography on Toyopearl HW-55F gave the values of 1.6×10^5 and 5.5×10^4 for the molecular weight of ginsenan PA and ginsenan PB, respectively. Quantitative analyses showed that
20 ginsenan PA contained 21.3% arabinose, 53.4% galactose, 2.0% rhamnose, 16.0% galacturonic acid and 2.7% glucuronic acid. The molar ratio of these component sugars was 11:22:1:6:1. Ginsenan PB contained 11.0% arabinose, 32.2% galactose, 8.1% rhamnose, 39.9% galacturonic acid, and 5.0% glucuronic acid. The molar ratio was
25 3:7:2:8:1. Both polysaccharides showed marked reticuloendothelial system-potentiating activity in a carbon clearance test, and pronounced anti-complementary activity and alkaline phosphatase-inducing activity in a dose dependent manner.

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In another study^[3], an additional two polysaccharides were isolated from the supernatant of the above extract treated with CTAB, i.e., the supernatant was poured into ethanol. The precipitate was separated and applied to columns of DEAE-Sephadex A-25 and
5 Sephadex G-25 to give another two pure polysaccharides, designated S-IA and S-IIA. Gel chromatography on Toyopearl HW-55F gave the values of 5.6×10^4 and 1.0×10^5 for the molecular weight of S-IA and S-IIA respectively. Ginsenan S-IA contains 42.3% arabinose, 50.8% galactose and 6.9% galacturonic acid with the molar ratio of 8:8:1.
10 Ginsenan S-IIA is composed of 42.0% L-arabinose, 32.6% galactose, 6.2% glucose, and 19.2% galacturonic acid. The molar ratio is 15:10:2:5.

Several polysaccharide fractions from leaves and roots of *Panax ginseng* have been separated by Yamada's group in the Oriental
15 Medicine Research Center of the Kitasato Institute, Japan. The chemical properties and biological activities were investigated and compared.^[4]

Ginseng roots and leaves from China, after treated with ethanol to remove their ginsenosides, were extracted with water, and the
20 residue were extracted with 0.5 M NaOH to give water-soluble and alkaline soluble polysaccharide fractions (designated as GR-2 and GL-2 for the water-soluble fractions and GRA-2 and GLA-2 for the alkaline-soluble fractions, respectively). Based on the acidity of the component polysaccharides, all fractions were further fractionated into strongly
25 acidic (designated as GR-3, GL-3, GRA-3, and GLA-3), weakly acidic (designated as GR-4, GL-4, GRA-4, and GLA-4), and neutral polysaccharide fractions (designated as GR-5, GL-5, GRA-5, and GLA-5), by treatment with cetyltrimethylammonium. The roots contained

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larger amount of polysaccharide than the leaves. The strongly acidic polysaccharide fractions from the roots had a high content of uronic acid, even higher than 50%. Similar component sugars were detected from all fractions. They were rhamnose, arabinose, galactose, glucose, galacturonic acid, and glucuronic acid. Galacturonic acid was the main uronic acid component.

The fraction with highest anti-complementary activity, GL-3, was further fractionated with columns of DEAE-Sephadex, Sepharose CL-6B, DEAE Toyopearl, and ethanol precipitation to give fractions designated as GL-PI through GL-PIV.^[5] All fractions contained 32-44% uronic acid. Fraction PI had the highest molecular weight of 50,000. PI and PII consisted mainly of Rha, Gal, and GalA, and PIII contained Fuc in addition, whereas PIV consisted of Gal, Glc, and GalA. A detailed structural determination was performed.

An anti-ulcer pectic polysaccharide (GL-BIII) was isolated from weakly acidic polysaccharide fraction GL-4 by chromatography on DEAE Sepharose CL-6B and Sepharose CL-6B. It was mainly composed of Rha, Ara, Man, Gal, Glc, GalA, and GlcA in the molar ratio of 3:4:2:10:1:7:4. Detailed structural determination was performed.^[6]

Another macrophage Fc receptor expression-enhancing polysaccharide (GL-4IIb2) was separated from GL-4 by anion-exchange chromatography on DEAE-Sepharose CL-6B. Chemical analysis showed that the sample contained 65% total carbohydrate and 33.7% uronic acid. The composition analysis and structural determination were performed.^[7]

Another *Panax ginseng* extract with anticomplementary activity, G-115, was studied by the same group.^[8] G-115 was fractionated in order to characterize the active substances for

anticomplementary and mitogenic activities. The most potent anticomplementary activity was observed in the crude polysaccharide fraction, G-115G, whereas the water-soluble dialyzable fraction, G-115E, showed the most potent mitogenic activity. G-115G was further
5 purified by precipitation with cetyltrimethylammonium bromide, anion-exchange chromatography on DEAE-Sepharose and gel filtration on Sepharose CL-4B, and a major potent anticomplementary polysaccharide, G-115I1-IIa-2-3 was obtained. This polysaccharide was homogeneous. Its molecular weight was estimated to be 3.68×10^5 . It
10 consisted mainly of arabinose, galactose and glucose in addition to small amounts of galacturonic acid, glucuronic acid and rhamnose.

Ginseng polysaccharides were isolated from Korean, Chinese, and Japanese ginseng by Hikino's group at the Pharmaceutical Institute, Tohoku University, Japan. The hypoglycemic
15 activities of the ginseng polysaccharides have been tested. The composition and some structure feature have been elucidated.^[9-14]

Three polysaccharides, quinquefolans A through C, were isolated from American ginseng.^[14] Their molecular weights were estimated to be higher than 2.0×10^6 by gel chromatography over
20 Sephacryl S-500. The neutral sugar components were mannose and glucose (molar ratio, 1.0:2.3) for quinquefolan A, mannose and glucose (1.0:5.5) for quinquefolan B, and xylose for quinquefolan C. The acidic sugar components in quinquefolans A through C were found to be 10.8, 11.7, and 7.1% respectively. The content of peptide moieties in these
25 glycans was 2.7, 2.9, and 2.3% for quinquefolans A through C respectively. All of these polysaccharides showed hypoglycemic effects in normal and alloxan-reduced hypoglycemic mice.

An acidic polysaccharide with the molecular weight of

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150,000, called ginsan, was isolated from *Panax ginseng* by a research group at the Laboratory of Immunology, Korean Cancer Center Hospital, Seoul, Korea.^[15] This polysaccharide was composed of 3.7% protein and 47.1% hexose (glucose and galactose) and 43.1% uronic acid (galacturonic acid). Ginsan induced the proliferation of T cells and B cells and generated lymphokine activated killer cells from both natural killer and T cells through endogenously produced multiple cytokines.^[16]

Miao et al. from Northeast Normal University of China has isolated polysaccharides from American ginseng. The purification and structural analysis were performed.^[17]

The biological activities of polysaccharides from American ginseng have been investigated by a research group in Norman Bethune University of Medical Science.^[18, 19] They found that polysaccharides from American ginseng (PPQ) enhanced lymphocyte transformation. The effect of polysaccharide from *Panax quinquefolium* (PPQ-1) on cytokine production from murine spleen lymphocyte *in vitro* was studied. The data suggest that PPQ-1 regulates immune function.

Summary of the Invention

The present inventors have found that certain American ginseng extracts have immunoregulating properties. CVT-E002, and purified fractions PQ₂ and PQ₂₂₃ therefrom, specifically stimulates murine spleen cells to proliferate B cells, which subsequently produce a large amount of antibody. The fractions also increase serum immunoglobulin (e.g., total IgG) levels and stimulate macrophages to produce IL-1, IL-6 and TNF- α . These fractions may be used for the prevention or treatment of general infection and other immune deficiency associated diseases.

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Therefore, the present invention is directed to processes of preparing ginseng fractions PQ₂, PQ₂₂₃ and CVT-E002 from samples of American ginseng.

Specifically, a process of preparing ginseng fraction PQ₂

5 comprises:

-combining American ginseng with a first solvent comprising an alcohol and heating the resulting solution at a temperature of about 80-100°C for a time period of about 2-4 hours to produce a first ginseng solution;

10 -thereafter separating the first ginseng solution to produce an alcohol/ginseng solution and a first ginseng residue;

-thereafter combining the first ginseng residue with water and heating the resulting solution at a temperature of about 80-100°C for a time period of about 2-4 hours to produce a ginseng residue
15 solution;

-thereafter separating the ginseng residue solution to produce a second ginseng residue and a first aqueous extract solution containing a first ginseng extract;

-providing a second aqueous extract solution which
20 comprises at least a part of the first ginseng extract, wherein in the second aqueous extract solution the proportion of the first ginseng extract to water is about 1:18 to 1:22;

-thereafter combining the second aqueous extract solution with a second solvent comprising an alcohol, wherein the
25 proportion of the second solvent to water is about 1:1 to 3:5, to produce a first precipitate and a first supernatant;

-thereafter combining the first supernatant produced in the previous step with a third solvent comprising an alcohol, wherein

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the proportion of the third solvent to first supernatant is about 3:2 to 3:1, to produce a second precipitate and a second supernatant; and

-isolating the second precipitate to produce ginseng fraction PQ₂.

5 A process of preparing ginseng fraction PQ₂₂₃ comprises:

-providing ginseng fraction PQ₂, as described above;

-fractionating the ginseng fraction PQ₂ to produce a first elution fraction and a second elution fraction, wherein the first elution fraction corresponds to a carbohydrate peak observed between
10 35 and 50 ml of elution volume and the second elution fraction corresponds to a carbohydrate peak observed between 50 and 85 ml of elution volume, as determined by gel filtration chromatography using the following materials:

(1) a chromatographic column containing a
15 matrix of a spherical cross-linked co-polymer of allyl dextran and N,N'-methylenebisacrylamide, having a bed dimension of 16 × 600 mm, a bed volume of 120 ml, and a fractionation range (MW) of 5000 to 250,000 for globular proteins and 1000 to 80,000 for dextrans, and

(2) an elution buffer of Tris-HCl containing
20 0.1 N HCl and 0.3 M NaCl at a pH of 7.0;and

-isolating and combining the first elution fraction and the second elution fraction to produce ginseng fraction PQ₂₂₃.

A process of preparing ginseng fraction CVT-E002 comprises:

25 -combining American ginseng with a first solvent comprising an alcohol in a proportion of about 7-9 ml of first solvent per gram of ginseng and heating the resulting solution at a temperature of about 80-100°C for a time period of about 2-4 hours, to produce a first

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ginseng solution;

-thereafter separating the first ginseng solution to produce an alcohol/ginseng solution and a first ginseng residue;

-thereafter combining the first ginseng residue with
5 water in a proportion of about 7-9 ml of water per gram of ginseng residue and heating the ginseng residue solution at a temperature of about 80-100°C for a time period of about 2-4 hours, to produce a ginseng residue solution;

-thereafter separating the ginseng residue solution to
10 produce a second ginseng residue and an aqueous extract solution containing a ginseng extract; and

-drying the aqueous extract solution to produce ginseng fraction CVT-E002.

The present invention also includes ginseng fractions PQ₂,
15 PQ₂₂₃ and CVT-E002, which are prepared according to the processes described above.

The invention further includes ginseng fractions having specific carbohydrate contents.

A first ginseng fraction has a carbohydrate content which
20 comprises about 2-6 mol% rhamnose, about 41-49 mol% galacturonic acid, about 12-18 mol% glucose, about 16-22 mol% galactose and about 12-19 mol% arabinose.

A second ginseng fraction has a carbohydrate content which comprises about 3-8 mol% rhamnose, about 36-44 mol%
25 galacturonic acid, about 2-7 mol% glucose, about 25-33 mol% galactose and about 17-25 mol% arabinose.

A third ginseng fraction has a carbohydrate content which comprises about 0.5-5 mol% rhamnose, about 11-22 mol% galacturonic

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acid, about 40-60 mol% glucose, about 10-19 mol% galactose and about 11-19 mol% arabinose.

The invention also includes pharmaceutical compositions, comprising the ginseng fractions of the invention, in combination with a pharmaceutically acceptable carrier.

The invention further includes the use of a ginseng fraction according to the invention, alone or in combination with another medicament, in the preparation of a pharmaceutical composition suitable for treating a condition characterized by low immunity.

The invention also includes the use of a ginseng fraction of the invention to stimulate the production of IL-1, IL-6 and/or TNF- α in cells.

The invention further includes the use of a ginseng fraction according to the invention to stimulate the *in vitro* or *in vivo* production of immunoglobulins.

Also included is the use of a ginseng fraction according to the invention to activate B-lymphocyte proliferation and antibody production therefrom.

The invention also includes a method of treating a condition characterized by low immunity in a patient in need thereof, comprising administering to the patient a condition treating effective amount of a ginseng fraction according to the invention.

Brief Description of the Drawings

Figure 1 shows a plot of elution volume against log molecular weight for standard dextran samples eluted on a chromatographic column.

Figure 2 shows a chromatogram of ginseng fraction CVT-E002.

Figure 3 shows a chromatogram of ginseng fraction PQ₁.

Figure 4 shows a chromatogram of ginseng fraction PQ₂.

Figure 5 shows a chromatogram of ginseng fraction PQ₃.

Figure 6 shows a chromatogram of ginseng fraction PQ₂₂₃.

Figure 7 shows the effect of ginseng fraction PQ₂₂₃ on mouse splenocyte
5 B cell proliferation.

Figure 8 shows the effect of ginseng fraction PQ₂₂₃ on mouse splenocyte
T cell proliferation.

Figure 9 shows the specificity of the effect of ginseng fraction PQ₂₂₃ on
mouse splenocyte T and B cell proliferation.

10 Figure 10 shows the effect of ginseng fraction PQ₂₂₃ on *in vitro* antibody
production by mouse spleen.

Figure 11 shows the enhancement of plaque forming cells (PFC) in mice
injected with ginseng fraction PQ₂₂₃.

Detailed Description of the Invention

15 A process of preparing ginseng fraction PQ₂ comprises:

(a) combining American ginseng with a first solvent
comprising an alcohol and heating the resulting solution at a
temperature of about 80-100°C for a time period of about 2-4 hours to
produce a first ginseng solution;

20 (b) thereafter separating the first ginseng solution to
produce an alcohol/ginseng solution and a first ginseng residue;

(c) thereafter combining the first ginseng residue with
water and heating the resulting solution at a temperature of about 80-
100°C for a time period of about 2-4 hours to produce a ginseng residue
25 solution;

(d) thereafter separating the ginseng residue solution to
produce a second ginseng residue and a first aqueous extract solution

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containing a first ginseng extract;

(e) providing a second aqueous extract solution which comprises at least a part of the first ginseng extract, wherein in the second aqueous extract solution the proportion of the first ginseng extract to water is about 1:18 to 1:22;

(f) thereafter combining the second aqueous extract solution with a second solvent comprising an alcohol, wherein the proportion of the second solvent to water is about 1:1 to 3:5, to produce a first precipitate and a first supernatant;

(g) thereafter combining the first supernatant produced in step (f) with a third solvent comprising an alcohol, wherein the proportion of the third solvent to first supernatant is about 3:2 to 3:1, to produce a second precipitate and a second supernatant; and

(h) isolating the second precipitate to produce ginseng fraction PQ₂.

The alcohol in each of the first solvent, second solvent and third solvent comprises an alcohol which is inert to the ginseng and is easily separated from the desired product. Those of skill in the art would readily be able to select alcohols which meet these requirements.

Preferably the alcohol in each case independently comprises a saturated or unsaturated C₁-C₆ alcohol. More preferably, the alcohol in each case independently comprises ethanol or methanol.

In each of steps (a) and (c), it is preferred that the resulting solution is heated for a time period of about 3 hours. It is also preferred that in step (a) the first solvent and the ginseng are combined in a proportion of about 7-9 ml of first solvent per gram of ginseng, most preferred about 8 ml of first solvent per gram of ginseng. In step (c), it is preferred that the water and the first ginseng residue are combined in

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a proportion of about 7-9 ml of water per gram of ginseng residue, most preferred about 8 ml of water per gram of ginseng residue.

Directly following step (d) in the process recited above, the first ginseng extract may optionally be concentrated. This may be accomplished in any way according to procedures well known to those of skill in the art. For example, the first aqueous solution containing the first ginseng extract may be centrifuged (e.g., at a speed of 2500 to 10,000 rpm for about 5-15 minutes). The first aqueous solution may also be filtered. Alternatively or in addition to concentrating the first ginseng extract, the first ginseng extract may be freeze dried for later use.

In step (e), the second aqueous extract solution comprises at least a part of the first ginseng extract. It is important that the ratio of first ginseng extract to water in the second aqueous extract solution be about 1:18 to 1:22, more preferably about 1:20. This may be achieved in any of a number of ways. If the first ginseng extract is concentrated and/or freeze dried following step (d), as described above, water should be added to the first ginseng extract to achieve the desired ratio. Alternatively, a part of the first aqueous extract solution, or the entire first aqueous extract solution, may be used in the second aqueous extract solution. If needed, additional water may be added to achieve the desired ratio. The use of at least a part of the first aqueous extract solution which contains the desired amount of first ginseng extract would avoid the need to conduct additional concentrating or freeze drying steps between steps (d) and (e).

In step (f), it is preferred that the proportion of the second solvent to water is about 3:4.

In step (g), it is preferred that the proportion of the third

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solvent to first supernatant is about 2:1.

A process of preparing ginseng fraction PQ_{223} comprises:

- (a) providing ginseng fraction PQ_2 , produced according to the process described above;
- 5 (b) fractionating the ginseng fraction PQ_2 to produce a first elution fraction and a second elution fraction, wherein the first elution fraction corresponds to a carbohydrate peak observed between 35 and 50 ml of elution volume and the second elution fraction corresponds to a carbohydrate peak observed between 50 and 85 ml of
10 elution volume, as determined by gel filtration chromatography using the following materials:
 - (1) a chromatographic column containing a matrix of a spherical cross-linked co-polymer of allyl dextran and N,N'-methylenebisacrylamide, having a bed dimension of 16 × 600 mm, a bed
15 volume of 120 ml, and a fractionation range (MW) of 5000 to 250,000 for globular proteins and 1000 to 80,000 for dextrans, and
 - (2) an elution buffer of Tris-HCl containing 0.1 N HCl and 0.3 M NaCl at a pH of 7.0; and
- (c) isolating and combining the first elution fraction and
20 the second elution fraction to produce ginseng fraction PQ_{223} .

The first elution fraction is also known as PQ_2A , and the second elution fraction is also known as PQ_2B . These elution fractions may be isolated separately. Additionally, the same process as noted above may be used to produce additional elution fractions PQ_2C (which
25 corresponds to a carbohydrate peak observed between 95 and 110 ml of elution volume, and PQ_2D (which corresponds to a carbohydrate peak observed between 120 and 250 ml of elution volume). Each of these fractions may also be separately isolated. In addition, compositions

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besides PQ₂₂₃ which comprise two or more of fractions PQ₂A through PQ₂D may be also made.

It is preferred that ginseng fraction PQ₂ is fractionated using gel filtration chromatography. However, any other types of fractionation
5 known to the skilled artisan are suitable.

The procedure for performing gel filtration chromatography is well known by those of ordinary skill in the art, following manufacturer's recommendations as to flow rate, sample volume and temperature at which the procedure should be performed. Variance of
10 these factors within the manufacturer's specifications does not significantly affect the results of the chromatographic run.

Determination of carbohydrate content may be made by any procedure known in the art. It is preferred that a microtiter plate assay be conducted for the determination of the carbohydrate composition of
15 the fraction. Such an assay is well known to those of skill in the art. See, for example, Dubois et al. *Anal. Chem.* **28**: 350-56 (1956), hereby incorporated by reference. Absorbance of the sample provided according to the microtiter plate assay is preferably conducted at 492 nm.

20 A process of preparing ginseng fraction CVT-E002 comprises:

(a) combining American ginseng with a first solvent comprising an alcohol in a proportion of about 7-9 ml of first solvent per gram of ginseng and heating the resulting solution at a temperature of
25 about 80-100°C for a time period of about 2-4 hours, to produce a first ginseng solution;

(b) thereafter separating the first ginseng solution to produce an alcohol/ginseng solution and a first ginseng residue;

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(c) thereafter combining the first ginseng residue with water in a proportion of about 7-9 ml of water per gram of ginseng residue and heating the ginseng residue solution at a temperature of about 80-100°C for a time period of about 2-4 hours, to produce a
5 ginseng residue solution;

(d) thereafter separating the ginseng residue solution to produce a second ginseng residue and an aqueous extract solution containing a ginseng extract; and

(e) drying or concentrating the aqueous extract solution
10 to produce ginseng fraction CVT-E002.

The alcohol in the first solvent comprises an alcohol which is inert to the ginseng and is easily separated from the desired product. Those of skill in the art would readily be able to select alcohols which meet these requirements. Preferably the alcohol comprises a saturated
15 or unsaturated C₁-C₆ alcohol. More preferably, the alcohol comprises ethanol or methanol.

It is preferred that in step (a) the first solvent and the sample are combined in a proportion of about 8 ml of first solvent per gram of sample. It is also preferred that in step (c) the water and the
20 first ginseng residue are combined in a proportion of about 8 ml of water per gram of ginseng residue.

In steps (a) and (c), it is preferred that the first ginseng solution is heated for a time period of about 3 hours.

The invention also includes several ginseng fractions.

25 A first ginseng fraction has a carbohydrate content which comprises about 2-6 mol% rhamnose, about 41-49 mol% galacturonic acid, about 12-18 mol% glucose, about 16-22 mol% galactose and about 12-19 mol% arabinose. Preferably, the carbohydrate content

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comprises about 3-5 mol% rhamnose, about 43-47 mol% galacturonic acid, about 14-16 mol% glucose, about 18-20 mol% galactose and about 14-17 mol% arabinose. Most preferably, the carbohydrate content comprises about 4 mol% rhamnose, about 45 mol% galacturonic acid,
5 about 15 mol% glucose, about 19 mol% galactose and about 15 mol% arabinose.

A second ginseng fraction in accordance with the invention has a carbohydrate content which comprises about 3-8 mol% rhamnose, about 36-44 mol% galacturonic acid, about 2-7 mol% glucose, about 25-
10 33 mol% galactose and about 17-25 mol% arabinose. Preferably, the carbohydrate content comprises about 4-7 mol% rhamnose, about 37-42 mol% galacturonic acid, about 3-6 mol% glucose, about 27-32 mol% galactose and about 19-24 mol% arabinose. Most preferably, the carbohydrate content comprises about 5 mol% rhamnose, about 39
15 mol% galacturonic acid, about 4 mol% glucose, about 29 mol% galactose and about 21 mol% arabinose.

A third ginseng fraction according to the invention has a carbohydrate content which comprises about 0.5-5 mol% rhamnose, about 11-22 mol% galacturonic acid, about 40-60 mol% glucose, about
20 10-19 mol% galactose and about 11-19 mol% arabinose. Preferably, the carbohydrate content comprises about 1-3 mol% rhamnose, about 13-20 mol% galacturonic acid, about 42-57 mol% glucose, about 12-17 mol% galactose and about 13-17 mol% arabinose.

The invention also includes pharmaceutical compositions,
25 comprising any of the ginseng fractions according to the invention in combination with a pharmaceutically acceptable carrier. Those of skill in the art are familiar with any pharmaceutically acceptable carrier which would be useful in this regard, and therefore the procedure for making

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pharmaceutical compositions in accordance with the invention will not be discussed in detail. Suitably, the pharmaceutical compositions may be in the form of tablets, capsules, liquids, lozenges, lotions or suppositories.

5 The invention includes the use of a ginseng fraction according to the invention in the preparation of a pharmaceutical composition suitable for treating a condition characterized by low immunity, such as common cold, influenza, chronic fatigue syndrome, AIDS and cancer. The ginseng fraction may be used alone or in
10 combination with another medicament. The ginseng fractions of the invention are especially suitable for co-administration with a chemotherapeutic agent or as a supplement to radiation therapy, since cancer patients are known to have serious suppression of the immune system.

15 The invention also includes a method of treating a condition characterized by low immunity in a patient in need thereof, comprising administering to the patient a condition treating effective amount of a ginseng fraction according to the invention. Preferably, the condition is selected from the group consisting of common cold, influenza, chronic
20 fatigue syndrome, AIDS and cancer. Dosages of ginseng fractions in accordance with the invention depend upon the particular condition to be treated, as well as the age, sex and general health condition of the patient. However, suitable dosages may be found in the range between 1 and 5000 mg/kg body weight per day, in between 1 and 10 daily
25 doses. The ginseng fractions may be administered orally, via injection or infusion, topically, nasally, ocularly, vaginally or rectally.

The invention will now be further elucidated by the following Examples.

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EXAMPLE 1: A First Process for Preparing
Fraction CVT-E002 and Purifying This Fraction

American ginseng root was chemically extracted and purified sequentially to give fractions CVT-E001 and CVT-E002. An amount of CVT-E002 was further purified to give fractions G₁, G₂ and G₃.
5 The detailed description of the procedure is as follows.

500 grams of dried ground root of ginseng was extracted with 4 liters of 85% ethanol or 3 liters of 90% methanol on a water bath at 80-85°C while stirring for 3 hours and filtered to give an alcohol
10 solution and residue. The alcohol solution was concentrated and spray dried to give a product of total saponins (CVT-E001). The residue was extracted with 4 liters of water on a water bath at 95-100°C while stirring
for 3 hours. The extract was filtered through a muslin bag and
centrifuged to give an aqueous solution containing CVT-E002, and the
15 remaining residue was discarded.

A portion of the aqueous solution was used for further purification, and an equal volume of 95% ethanol was added to the aqueous solution, which caused precipitation. The precipitate was centrifuged and lyophilized to give the fraction G₁. The supernatant was
20 reduced in volume by evaporation. An equal volume of 95% ethanol was added to give the next precipitate, G₂, and the remaining supernatant was removed with ethanol and lyophilized to produce a powder, G₃.

The further purification of G₂ was conducted as follows.
25 Two grams of G₂ were dissolved in 80 ml of water and dialyzed against 3 volumes of water in a Sigma D-7884 dialysis tubing with a 1200 molecular weight cut-off. The dialysis was done at 4°C for 72 hours, collecting dialysate twice every 24 hours. The resulting dialysate was

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concentrated to 15 ml, then precipitated with an equal volume of methanol. The precipitate was dissolved in water and lyophilized to yield a powder, G₂₂.

The further purification of G₂₂ was conducted as follows.

- 5 1.5 grams of G₂₂ were dissolved in 60 ml water and then dialyzed against 600 mls of water in a Fisher Spectral/Por molecularporous membrane tubing with a 1000 molecular weight cut-off at 4°C. The first dialysate collected after 24 hours was concentrated to 15 mls using an evaporator and then lyophilized. The resulting dried powder was
- 10 referred to as G₂₂₁. In a similar manner, a second batch of dialysate was collected after 48 hours and referred to as G₂₂₂. The retentate was concentrated and lyophilized to give a powder called G₂₂₃.

**EXAMPLE 2: A Second Process for Preparing
Fraction CVT-E002 and Purifying This Fraction**

- 15 An alternative process for preparing ginseng fraction CVT-E002 according to the invention is as follows.

- 1000 grams of dried ground root of American ginseng was extracted with 8 liters of 85% ethanol on a water bath at 95-100°C while stirring for 3 hours and filtered to give an alcohol solution and residue.
- 20 The residue was combined with water (1:8) on a hot water bath with continuing agitation for 3 hours. After cooling to room temperature, the mixture was filtered. The filtrate was centrifuged at 5000 rpm for 10 min. The supernatant was concentrated and freeze dried to give extract CVT-E002. The amount of CVT-E002 produced by this method is
- 25 approximately the same as produced according to Example 1, i.e., about 10% of the weight of the original raw ginseng.

A portion of CVT-E002 was further fractionated as follows.

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1600 ml of 95% ethanol was added to a solution of 100 grams of CVT-E002 powder in 2000 ml of water. The precipitate was isolated as PQ₁. The supernatant was concentrated to 500 ml. Another portion of 95% ethanol (1000 ml) was added to this concentrated solution again to give
5 second precipitate fraction. The precipitate was isolated and freeze dried to give fraction PQ₂. The supernatant was concentrated and freeze dried to give PQ₃.

EXAMPLE 3: Mitogenic Activity Test

Different fractions of American ginseng at various levels of
10 purification were selected by screening on the basis of their mitogenic activity of lymphocytes *in vitro*. Though mitogenicity is considered a rather artificial event in relation to the normal events occurring in the immune system *in vivo*, mitogens provide good indications of possible effector function.

15 The test method for mitogenic activity is described as follows: Balb/C or C57B1/6J mice were used for the test. Balb/C mice were obtained from the Health Sciences Laboratory Animal Services facility (University of Alberta, Edmonton, Canada). C57B1/6J mice were obtained from the Jackson Laboratory (Bar Harbor, ME). Mice from 7
20 to 10 weeks old were age and sex matched for each experiment. Mice were killed by cervical dislocation. Spleens were removed by using aseptic techniques and were crushed between the frosted ends of two glass slides. After washing by centrifugation in Hanks Balanced Salt Solution (HBSS), the cells were suspended in RPMI 1640 medium pH
25 7.4 (Gibco, Grand Island, N.Y.) containing 10% fetal bovine serum (Flow Labs), 50 mM mercaptoethanol (ICN Pharmaceuticals, Plainview, N.Y.), and penicillin-streptomycin (Gibco). Cultures were set up in triplicate in

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96-well flat-bottomed Linbro plates at 1.25×10^6 cells per ml final concentration. Experimental groups were set up with testing fractions previously filter-sterilized and dissolved in HBSS. Control cultures consisted of a group without mitogen and groups with 20 $\mu\text{g/ml}$ phytohemagglutinin (PHA) or 25 $\mu\text{g/ml}$ lipopolysaccharide (LPS). It is known that PHA specifically stimulates the T- type of spleen cells while LPS stimulates the B-type of spleen cells. The cultures were incubated at 37°C in a humidified 5% CO_2 atmosphere for 72 hours. Four hours before the end of the 72 hour incubation, 1 μCi of tritiated thymidine (New England Nuclear, Boston, Massachusetts) was added to each well. The cells were harvested with an automated sample harvester (Skatron, Virginia) and then incorporated radioactivity was assayed by scintillation counting. The results were calculated as a % of control (mean \pm standard deviation, in triplicate).

The different extraction conditions shown in Examples 1 and 2, such as concentration of CVT-E002 in water and the volume of ethanol used for precipitation, affect the further fractionation of the CVT-E002. The products obtained from these two processes were compared for mitogenic activity. The results are shown in the following Tables. Each test was performed in triplicate with three batches.

Table 1. Mitogenic activity of G_1 , G_2 , and G_3 obtained from procedure according to Example 1

Fractions	Activity (% control)		
	Batch 1	Batch 2	Batch 3
G_1	4090 ± 1644	2845 ± 1318	4446 ± 977
G_2	3015 ± 1284	4101 ± 1671	3146 ± 318

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G ₃	1791 ± 573	1735 ± 96	365 ± 26
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Table 2. Mitogenic activity of PQ₁, PQ₂, and PQ₃ obtained from procedure according to Example 2

Fractions	Activity (% control)		
	Batch 1	Batch 2	Batch 3
PQ ₁	2448 ± 454	3169 ± 431	2492 ± 138
PQ ₂	6427 ± 1609	6467 ± 1593	7034 ± 1834
PQ ₃	1313 ± 288	1522 ± 342	1622 ± 222

As Tables 1 and 2 indicate, the change in extraction procedure between Examples 1 and 2 correlates with a transfer of biological activity from the G₁ fraction to the PQ₂ fraction.

EXAMPLE 4: Fractionation of PQ₂ and a Process for Preparing Fraction PQ₂₂₃

CVT-E002, PQ₁, PQ₂ and PQ₃ according to Example 2 were further fractionated according to their molecular weight distribution by gel filtration chromatography over a HiPrep 16/60 Sephacryl S-200 high resolution column (Pharmacia Biotech, Cat. No. 17-1166-01), which contains a matrix of a spherical cross-linked co-polymer of allyl dextran and N,N'-methylenebisacrylamide, has a bed dimension of 16 × 600 mm, a bed volume of 120 ml, and a fractionation range (MW) of 5000 to 250,000 for globular proteins and 1000 to 80,000 for dextrans. Dextran samples (MW=71.4k, 37.5k, 19.5k and 9.5k) were purchased from Sigma and were used as standard samples. 5 mg of each sample was dissolved in 1 ml of water, loaded onto the column, and eluted with Tris-

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HCl buffer containing 0.1 N HCl and 0.3 M NaCl, pH 7.0, flow rate 0.3 ml/min and environmental temperature 4°C. A volume of eluate was collected as a number of individual 5 ml portions.

Total carbohydrate content of each individual portion of eluate was tested with a microtiter plate assay for total carbohydrate content. This is a modification of a method described in Dubois et al. *Anal. Chem.* **28**: 350-56 (1956). The total carbohydrates in the eluate are derivatized in order to be detectable at a certain absorbance spectrum. D-glucose was used as the standard sample, and other materials used were concentrated sulfuric acid (98%) and 5% phenol. The polystyrene microtiter plates were purchased from SARSTDT (Quebec, Canada). To each well of a microtiter plate, 40 µl of a standard sample solution containing 1 to 10 µg of D-glucose was applied and 40 µl of 5% phenol was added and mixed. 200 µl of concentrated sulfuric acid was then carefully added. After mixing reagent and individual portion of eluate with a multichannel pipette, the plate was incubated under 80°C for 1 hour. After cooling to room temperature, the absorbance of the sample was measured at 492 nm on a Multiskan microtiter plate reader. The data was saved and the gel filtration chromatography results were plotted using the Microsoft Excel program.

A standard curve was created by running a series of standard dextrans (Sigma) with known molecular weights over the same column (Figure 1). The chromatograms for CVT-E002 and lots 21 of PQ₁, PQ₂ and PQ₃ are shown in Figures 2 -5. In each Figure, points on the x-axis show the elution volume in terms of one-fifth of the actual volume. In order to get a true value of elution volume, therefore, the results in the Figures must be multiplied by five.

The gel filtration chromatogram of CVT-E002 (Figure 2)

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showed mainly three peaks. The molecular weight for first peak (elution fraction 7-10) was estimated to be 70,000 or higher according to the standard curve (Figure 1). The molecular weight for the third peak (elution fraction 18-22) was estimated to be about 1,000. A small broad second peak (elution fraction 10-17) was also observed. The ratio of these three peaks was 57.2:8.5:34.4.

The chromatogram of PQ_1 (Figure 3) showed one main peak (elution fraction 7-10), which corresponds to the high molecular weight fraction of CVT-E002, and a minor peak (elution fraction 18-21). The ratio of the two peaks was 86.8:13.2.

The chromatogram of PQ_2 (Figure 4) showed three main peaks. The peak designated A (elution fraction 7-10) and the peak designated C (elution fraction 19-22) correspond to the two main peaks of CVT-E002. Another broad peak, designated B (elution fraction 10-17), was observed between peaks A and C. The molecular weight for this peak was estimated to be about 2,000-60,000. The ratio of the three peaks A-C was 34.1:45.7:20.3. A region of small peaks, designated D (elution fraction 24-50, a portion of which is shown in Figure 4), was also observed.

The chromatograph of PQ_3 (Figure 5) showed only one main peak (elution fraction 18-21), which corresponded to the low molecular weight fraction of CVT-E002.

Fractions A, B, C and D of PQ_2 were collected and freeze dried. The dried powders were renamed as PQ_2A , PQ_2B , PQ_2C and PQ_2D , respectively. Fractions A and B were also combined, and the combined fraction was called PQ_{223} (a chromatograph of which is shown in Figure 6). The mitogenic activity of PQ_2A , PQ_2B , PQ_2C and PQ_2D were tested and compared with the activity of fractions PQ_{223} , PQ_2 and

CVT-E002. The results are shown in the following Table.

Table 3. Mitogenic activity of American ginseng fractions
(³H-thymidine incorporation as % of control)

5	Sample	Dose (µg/ml)	Activity (% of Control)
	Control	n/a	100
	PHA	20	1334.5 ± 158*
	LPS	25	3227 ± 177**
	PQ ₂ A	100	3002 ± 147**
10	PQ ₂ B	100	947 ± 42**
	PQ ₂ C	100	186.5 ± 10.5*
	PQ ₂ D	100	201 ± 5**
	CVT-E002	100	1395.5 ± 159.5*
	PQ ₂	100	2321.5 ± 8.5**
15	PQ ₂₂₃	100	3237 ± 13**

* and ** represent $P < 0.05$ and $P < 0.01$, respectively.

n = 3.

As the results indicate, the order of the potency is PQ₂₂₃ > PQ₂ > CVT-E002.

20 EXAMPLE 5: Chemical Determination of Total Carbohydrate and Protein Composition of CVT-E002, PQ₁, PQ₂, PQ₃, and PQ₂₂₃

Three batches of each fraction CVT-E002, PQ₁, PQ₂, PQ₃, and PQ₂₂₃ were obtained, and the total carbohydrate and protein composition were measured in each fraction. The total carbohydrate content was determined by the phenol-sulfuric method, as known in the art. The protein content was assayed by Lowry method, as known in the art, and bovine serum albumin was used as standard. The results are

25

shown in the following Table.

Table 4. Comparison of Protein and Carbohydrate content of fractions

	Sample	Protein Content (%)	Total Carbohydrate (%)
5	E002-1	9.32	66.4
	E002-2	7.38	69.9
	E002-5	8.31	60
	PQ ₁ -24	2.6	90.3
	PQ ₁ -25	2.6	93.6
10	PQ ₁ -27	2.9	96.1
	PQ ₂ -24	5.6	52.6
	PQ ₂ -25	5.4	46.8
	PQ ₂ -27	5.3	43.6
	PQ ₃ -24	7.9	52.6
15	PQ ₃ -25	8.5	64.2
	PQ ₃ -27	7.1	68.0
	PQ ₂₂₃ -7	5.8	78.0
	PQ ₂₂₃ -8	2.8	81.0
	PQ ₂₂₃ -9	2.9	85.2

20

EXAMPLE 6: Chemical Determination of Monosaccharide

Composition of CVT-E002, PQ₁, PQ₂ and PQ₂₂₃

Since total carbohydrate content mainly represented the amount of polysaccharides and oligosaccharides, the molar ratios of monosaccharides for certain fractions were also determined.

25

Lots of fractions CVT-E002, PQ₁, PQ₂ and PQ₂₂₃ were subjected to acid catalyzed hydrolysis to liberate their structural units,

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monosaccharides. A qualitative and quantitative analysis of the liberated monosaccharides was performed by HPLC after they were derivatized with MPP (3-methyl-1-phenyl-2-pyrazolin-5-one). The detailed methods are described as follows: a sample solution in 2 N HCl

5 was heated at 95-100°C for 6 h. The mixture was neutralized with NaOH solution. An internal standard was added. MeOH and aqueous NaOH solution (0.3 N) were then added to this mixture. This was then stirred at 50-60°C for 8 hr (or at room temperature for one week). The mixture was diluted (1:10) with water and analyzed by HPLC. The MPP-

10 derivatized samples were analyzed with a C-18 column, with an eluent of 18% acetonitrile in 0.1 M phosphate buffer (pH 7), at a flow rate of 1 ml/min, UV detector at 245 nm. The following Table shows the molar ratio of monosaccharides of CVT-E002, PQ₁, PQ₂ and PQ₂₂₃. Trace amounts of glucuronic acid was found in some CVT-E002, PQ₁ and PQ₂

15 samples.

Table 5. Monosaccharide Composition of CVT-E002, PQ₁, PQ₂ and PQ₂₂₃ (% in molar)

Sample	Rhamnose	Galacturonic Acid	Glucose	Galactose	Arabinose
E002-1	2.8	17.9	53	13.3	13
20 E002-2	1	13.1	56.7	12.7	15.2
E002-3	2.9	20.4	42.1	16.5	16.3
PQ ₁ -21	0.67	8.27	82.6	4.57	3.72
PQ ₁ -22	0.48	6.38	85	4.56	3.55
PQ ₁ -23	0.48	8.24	80.7	4.68	4.72
25 PQ ₂ -22	3.6	45.1	15.4	19.8	16.2
PQ ₂ -26	3.8	44.2	15.6	19	14.4

	PQ ₂ -27	3.8	44.2	15.6	19	14.4
	PQ ₂ -28	4	45.3	13.8	20.5	16.3
	PQ ₂₂₃ -7	5.5	40.8	4.1	28.2	20.3
	PQ ₂₂₃ -8	5.4	38.1	3.9	29.5	22.1
5	PQ ₂₂₃ -9	5.4	36.7	5.4	28.8	22.7

As the result indicate, the order of glucose content is as follows: PQ1 > CVT-E002 > PQ2 > PQ3; and the content of the galacturonic acid is as follows: PQ2 > PQ223 > CVT-E002 > PQ1.

EXAMPLE 7: Immune Regulation of Ginseng Fractions

10 1. **Macrophage activity**

The effects of CVT-E002, PQ₂, G₂ and PQ₂₂₃ on murine macrophages were studied *in vitro* using peritoneal exudate macrophages from C57B1/6 mice which preferentially mount a cell-mediated response and Balb/c mice which preferentially mount an antibody-mediated response. IL-1, IL-6 and TNF- α production were measured after macrophages were stimulated with 100 μ g/ml of test samples. The methods are described as follows.

A. Preparation of macrophages from peritoneal exudate cells and cell supernatant.

- 20 1). 1 ml of 3% thioglycollate was injected into the murine peritoneal cavity and peritoneal exudate macrophages were harvested from the peritoneal cavity after 3 days.
- 2). Cell suspension was washed twice with Hanks buffer by centrifugation at 1100 rpm at 4°C for 5 min.
- 25 3). Macrophages were suspended in RPMI-10% FBS medium.
- 4). Cells were counted and diluted to a final concentration of 10⁶/ml with

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RPMI-10% FBC medium.

5). Macrophages were cultured in $10 \times 10^6/10\text{ml}$ RPMI-10%FBS at 37°C for 2 hrs. Supernatant was discarded and the precipitated macrophages were washed with 10 ml of PBS twice.

- 5 6). Macrophages were harvested and re-suspended in RPMI-10% FBS.
7). Macrophage (5×10^5) were cultured with 10 mg/ml of LPS or 100 mg/ml of test sample for 24 or 48 hrs, after which time the supernatants were collected and filter sterilized for the bioassay (IL-1, IL-6 and $\text{TNF-}\alpha$). Experiments were carried out in triplicate.

10 B. IL-1 determination in supernatant.

The NOB1 cell line is a TK-variant of the EL4 cell line that generates CTLL growth stimulating activity (IL-2) in response to IL-1. Since NOB1 cells do not incorporate $^3\text{HTdR}$, a response to IL-1 can be measured in terms of uptake by the CTLL line. The procedure is
15 described as follows.

- 1). 10^4 CTLL cells were combined with 4×10^4 NOB1 cells in 200 μl of 5% FBS in IMDM in each well of a 96-well flat-bottom microtiter plate. Serially diluted IL-1 test samples of 4,8,16 or 32-fold or serial dilutions of IL-1 standards were applied. The final volume in each well was 200
20 μl . Each sample was prepared in triplicate.
2). Plates were incubated for 24 hr in a 37°C 5% CO_2 humidified incubator.
3). [^3H]-thymidine was added for the last 5 hr of incubation.
4). Cells were harvested and [^3H]-thymidine incorporation was
25 measured by liquid scintillation counting.
5). IL-1 concentration was calculated. The results are shown in the following Table.

Table 6. Effect of ginseng fractions on IL-1 production by murine macrophages

Macrophages ¹ stimulated with. . .	Production of IL-1 ² (pg/ml)
cells only	15.7 ± 1.0
5 LPS	27.7 ± 1.9**
CVT-E002	18.1 ± 1.2*
PQ ₂	19.5 ± 0.9*
G ₂	17.7 ± 0.7*
10 PQ ₂₂₃	19.9 ± 1.4*

1. Peritoneal exudate macrophages 5×10^5 /ml were cultured with 100 µg/ml samples, 10 µg/ml LPS for 48 hrs and culture supernatant was harvested.

2. Assay of IL-1 was performed using NOB1 and CTLL cell lines and results were expressed as mean ± SD

* P < 0.05; ** P < 0.01

15 The results show that CVT-E002, PQ₂, G₂ and PQ₂₂₃ significantly stimulated macrophages from C57B1/6 mice to produce IL-1. All test samples showed a similar potency of stimulating IL-1 production.

C. Determination of IL-6

20 The proliferation of a murine B cell hybridoma cell line, B9, is IL-6-dependent. B9 cells were cultured in a series of microwells containing decreasing concentration of the test samples. The procedure was as follows.

- 1). The number of B9 cells in an aliquot removed from the stock culture flask was counted at their log-phase of growth.
- 25 2). The cells were centrifuged for 5 min in a tabletop centrifuge at 180 x g, 4°C, and the pellet was resuspended in 10 ml of complete RPMI-10 medium. The procedure was repeated twice and the cells were resuspended in complete RPMI-10 medium at 2×10^3 cells/ml.
- 3). 100 µl of washed cells (2×10^3 cells/ml) was added to each well of

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a 96-well microtiter plate.

4). 100 μ l of 2-fold serial dilutions of the test samples were added to each well, reserving two or three rows of the plate for IL-6 standards.

5). The plates were incubated for 72 hr in a humidified 37°C, 5% CO₂ incubator.

6). [³H]-thymidine was added and plates were incubated for 4 hr at 37°C.

7). The cells were harvested and [³H]-thymidine incorporation was determined using a liquid scintillation counter.

8). IL-6 concentration was calculated. The results are as shown in the following Table.

Table 7. Effect of ginseng on IL-6 production by murine macrophages

Macrophages ¹ stimulated with. . .	Production of IL-6 ² (pg/ml)	
	C57B1/6 mice	Balb/c mice
cells only	71.7 \pm 7	102 \pm 1.8
LPS	14312.5 \pm 269.4**	22496.8 \pm 2381.2**
CVT-E002	181.5 \pm 2.1**	185.5 \pm 10*
PQ ₂	219.3 \pm 30.3**	243.4 \pm 6.4**
PQ ₂₂₃	2058.3 \pm 137.9**	2387.2 \pm 301.6**

1. Peritoneal exudate macrophages 5 x 10⁵/ml were cultured with 100 μ g/ml samples, 10 μ g/ml LPS for 24 hrs and culture supernatant was harvested.

2. Assay of IL-6 was performed using B9 cell line and results were expressed as mean \pm SD

* P < 0.05; ** P < 0.01

CVT-E002, PQ₂ and PQ₂₂₃ increased the production of IL-6 in supernatant of macrophages from C57B1/6 and Balb/c mice. The potency is PQ₂₂₃ > PQ₂ > CVT-E002.

D. Measurement of TNF- α

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The following protocol employed TNF-sensitive, actinomycin D-treated murine L929 fibroblasts to quantify TNF activity in supernatants derived from cell cultures. 1). 4×10^4 L929 cells in 50 μ l of IMDM-5% FSC were added into each well.

- 5 2). 50 μ l of test sample was added to the second well of each row (column 2). Two-fold serial dilutions were made by gently mixing the contents of well 2 and transferring 50 μ l from well 2 into well 3. The contents of well 3 were mixed gently, and 50 μ l from well 3 was transferred into well 4. This procedure was continued through well 12.
- 10 Finally, the contents of all wells in column 12 were gently mixed and 50 μ l was discarded from each well. At this point, all wells contained 50 μ l.
- 3). 50 μ l of actinomycin D solution was added to each well (2 μ l ml).
- 4). The plates were incubated for 24 hr at 37°C in 5% CO₂ in air.
- 5). 5 μ l of Neutral Red was added to each well and the plates were
- 15 incubated for 2.5 hours.
- 6). All supernatants in each well were quickly and carefully rinsed and emptied, and plates were washed with 200 μ l of PBS twice.
- 7). The PBS was aspirated and 100 μ l of 50% ethanol in 0.05M NaH₂PO₄ was added to each well and shaken for 5 min at room
- 20 temperature.
- 8). Each well was read immediately with a microtiter plate reader at an absorbance of 570 nm.
- 9). TNF- α concentration was calculated. The results are shown in the following Table.

25 Table 8. Effect of ginseng on TNF- α production by murine macrophages

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Macrophages ¹ stimulated with. . .	Production of TNF- α ² (pg/ml)	
	C57B1/6 mice	Balb/c mice
cells only	6.1 \pm 1.9	4.8 \pm 0.7
LPS	55.2 \pm 1.6**	71.4 \pm 15.5**
5 CVT-E002	19.7 \pm 2.5**	17.9 \pm 2.8**
PQ ₂	3.5 \pm 0.8	3.9 \pm 0.2
G ₂	6.1 \pm 1.2	7.2 \pm 0.1*
PQ ₂₂₃	16.9 \pm 2.5**	15.2 \pm 2.2**

- 10 1. Peritoneal exudate macrophages 5×10^5 /ml were cultured with 100 μ g/ml samples, 10 μ g/ml LPS for 48 hrs and culture supernatant was harvested.
 2. Assay of TNF- α was performed using L929-8 cell line and results were expressed as mean \pm SD of TNF- α production by macrophages.
 * P < 0.05; ** P < 0.01

15 TNF- α production was induced by CVT-E002 and PQ₂₂₃ in supernatant of macrophages from Balb/c or C57B1/6 cell lines. TNF- α in the supernatant of macrophages from Balb/c mice was significantly stimulated by G₂. PQ₂ seemed not to stimulate TNF- α production by macrophages.

2. Serum immunoglobulin production

20 Serum immunoglobulin production (total IgG) increased 21% in mice fed with CVT-E002, 26% in mice fed with PQ₂ and 31% in mice fed with PQ₂₂₃ compared with mice fed with water. The potency is PQ₂₂₃ > PQ₂ > CVT-E002. Results are shown in Table 9.

Table 10. *In vivo* immunoglobulin production by mice (n=15)

25 Mice fed with. . .	IgG (μ g/ml) \pm SD
	Water
	275.16 \pm 32.34
CVT-E002	333.01 \pm 50.88**

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PQ ₂	346.81 ± 31.39**
PQ ₂₂₃	400.46 ± 54.48**

3. Specificity of mitogenic activity of PQ₂₂₃

Well known mitogens such as PHA or LPS show specificity as to the type of cells they stimulate. It is important to know which specific type of spleen cells are stimulated by ginseng fractions. We therefore separated T cells by an affinity column and enriched for B cells by T cell depletion.

A. T-Cell enrichment

A single cell suspension of spleen cells was layered on lymphocyte-M (Cedarlane Labs) to remove red blood cells. The lymphocytes were then passed through an affinity chromatography column (Biotex Co. Ltd., Edmonton, Alberta) which removes B cells by adherence to the mouse immunoglobulin coated beads. The eluant consisted of an enriched population of T cells. Determination of the percentage of T cells relative to other cell types was done by the viability test using anti-thy 1.2 monoclonal antibody and complement treatment.

B. B-cell enrichment

After removal of red blood cells from the spleen cell population, the lymphocytes were incubated with monoclonal anti-thy 1.2 antibody for one hour at 37°C. Low-tox rabbit complement was then added and incubated for half an hour at 4°C. This procedure depleted the cell suspension of T cells, resulting in an enriched B cell population.

C. Culturing with PQ₂₂₃

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B or T cells were cultured with varying doses of PQ₂₂₃, and the responses were measured in terms of tritiated thymidine incorporation. Figures 7 and 8 show the proliferation effect of PQ₂₂₃ on relatively purified B and T cells, respectively. Control groups included cells
5 treated with LPS, a B cell-specific mitogen, and PHA, a T cell-specific mitogen. PQ₂₂₃ was found to be B cell-specific (see Figure 9).

4. The effect of PQ₂₂₃ on antibody production *in vitro*

Spleen cells were cultured for 72 hours in the presence of varying doses of PQ₂₂₃. Control cultures consisted of a group to which 25 µg of
10 LPS was added, and another to which no mitogen was added. The supernatants were tested for the presence of soluble immunoglobulins using the Enzyme Linked Immunosorbent Assay (ELISA). The supernatant containing soluble immunoglobulins was serially diluted in 0.1M tris buffer (pH 9). Microtiter plates were coated with this
15 supernatant and incubated at 4°C overnight. The plates were washed with PBS Tween three times, then 100 µl of the antibody-enzyme conjugate Goat F(ab')₂ anti-mouse Ig horseradish peroxidase (Tago Inc., Burlingame California) was added at a 1:1000 dilution. The plate was incubated for one hour at room temperature. The plate was again
20 washed three times with PBS, then 100 µl of ABTS peroxidase substrate (Kirkegaard and Perry Lab Inc.) was added to each well. One hour later absorbance was measured at a wavelength of 405 nm by a Flow Multiscan ELISA reader.

Figure 10 shows that 50 µg/ml PQ₂₂₃ stimulated the production of
25 immunoglobulins comparable to that of 25 µg LPS. A higher concentration of PQ₂₂₃, 500 µg/ml, did not elicit a higher degree of antibody production. This dose response curve is consistent with the

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proliferative responses of heterogeneous spleen cell population and purified B cells. These results were based on four experiments.

5. In vivo antibody plaque forming cells

Three concentrations of PQ₂₂₃ were i.v injected to three groups of 3 mice each. A control group was injected with a balanced salt solution. At the end of seven days of intravenous treatments with PQ₂₂₃, the mice were immunized with 0.2 ml of a 10% SRBC suspension in HBSS. Five days after immunization the mice were sacrificed and single cell suspensions from the spleens were prepared and adjusted to 4×10^6 cells per ml. A 0.1 ml aliquot of this cell suspension was mixed with 0.3 ml of a 10% SRBC solution guinea pig complement and 0.5 ml RPMI medium. Seventy microliters of this mixture was transferred to a Cunningham chamber. The chamber was sealed with paraplast wax and was incubated at 37°C for one hour prior to counting of plaque forming cells. All PFC levels are expressed as per one million spleen cells. Figure 11 shows that all three doses significantly enhanced antibody production in mice injected with the compound.

In summary, the results of the present studies indicate that the ginseng fractions of the invention activate macrophages to produce cytokines such as IL-1, IL-6 and TNF- α . They also activate lymphocytes, particularly B-lymphocyte proliferation and antibody production. The overall humoral mediated immune system was stimulated indicating the preventive effects on infections. The reported action of TNF- α production includes antiviral and anti-tumor benefits. TNF- α has also been reported to be of therapeutic benefit in the treatment of a variety of parasitic infections.

The following citations referred to in the Background of the

Invention are hereby incorporated by reference.

1. Tomoda et al. *Biol. Pharm. Bull.*, **16**, 22-5 (1993).
2. Tomoda et al. *Biol. Pharm. Bull.*, **17**, 1287-91 (1994).
3. Tomoda et al. *Biol. Pharm. Bull.*, **16**, 1087-90 (1993).
- 5 4. Gao et al. *Planta Medica*, **55**, 9-12 (1989).
5. Gao et al. *Carbohydr. Res.*, **181**, 175-87 (1988).
6. Kiyohara et al. *Carbohydr. Res.*, **263**, 89-101 (1994).
7. Shin et al. *Carbohydr. Res.*, **300**, 239-49 (1997).
8. Yamada et al. *Phytotherapy Res*, **9**, 264-9 (1995).
- 10 9. Konno et al. *Planta Medica*, **50**, 434-6 (1984).
10. Hikino et al., unpublished.
11. Oshima et al. *J. Ethnopharmacology*, **14**, 255-9 (1985).
12. Konno et al. *Int. J. Crude Drug Res.*, **25**, 53-6 (1987).
13. Konno et al. *J. Ethnopharmacology*, **14**, 69-74 (1985).
- 15 14. Oshima et al. *J. Natural Products*, **50**, 188-90 (1987).
15. Lee et al. *Anticancer Res.*, **17**, 323-32 (1997).
16. Kim et al. *Planta Medica*, **64**, 110-5 (1998).
17. Miao et al. *Shengwu Huaxue ZaZhi*, **9**, 610-4 (1993).
18. Ma et al. *Baiqiuen Yike Daxue Xuebao*, **23**, 236-8 (1997).
- 20 19. Zhu et al. *Zhongguo Yaolixue Tongbao*, **13**, 76-8 (1997).

We claim:

1. A process of preparing ginseng fraction PQ₂, the process comprising:

- 5 (a) combining American ginseng with a first solvent comprising an alcohol and heating the resulting solution at a temperature of about 80-100°C for a time period of about 2-4 hours to produce a first ginseng solution;
- (b) thereafter separating the first ginseng solution to produce an alcohol/ginseng solution and a first ginseng residue;
- 10 (c) thereafter combining the first ginseng residue with water and heating the resulting solution at a temperature of about 80-100°C for a time period of about 2-4 hours to produce a ginseng residue solution;
- (d) thereafter separating the ginseng residue solution to produce a second ginseng residue and a first aqueous extract solution
- 15 containing a first ginseng extract;
- (e) providing a second aqueous extract solution which comprises at least a part of the first ginseng extract, wherein in the second aqueous extract solution the proportion of the first ginseng extract to water is about 1:18 to 1:22;
- 20 (f) thereafter combining the second aqueous extract solution with a second solvent comprising an alcohol, wherein the proportion of the second solvent to water is about 1:1 to 3:5, to produce a first precipitate and a first supernatant;
- (g) thereafter combining the first supernatant produced in step
- 25 (f) with a third solvent comprising an alcohol, wherein the proportion of the third solvent to first supernatant is about 3:2 to 3:1, to produce a second precipitate and a second supernatant; and
- (h) isolating the second precipitate to produce ginseng fraction

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PQ₂.

2. The process of claim 1, wherein the alcohol in each of the first solvent, second solvent and third solvent independently comprises a saturated or unsaturated C₁-C₆ alcohol.
- 5 3. The process of claim 1, wherein the alcohol in each of the first solvent, second solvent and third solvent independently comprises ethanol or methanol.
4. The process of claim 1, wherein in step (e) the second aqueous extract solution comprises at least a part of the first aqueous extract
10 solution.
5. The process of claim 1, wherein in step (a) the resulting solution is heated for a time period of about 3 hours.
6. The process of claim 1, wherein in step (c) the resulting solution is heated for a time period of about 3 hours.
- 15 7. The process of claim 1, wherein in step (e) the proportion of the first ginseng extract to water is about 1:20.
8. The process of claim 1, wherein in step (f) the proportion of the second solvent to water is about 3:4.
9. The process of claim 1, wherein in step (g) the proportion of the
20 third solvent to first supernatant is about 2:1.

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10. The process of claim 1, wherein in step (a) the first solvent and the ginseng are combined in a proportion of about 7-9 ml of first solvent per gram of ginseng.
11. The process of claim 1, wherein in step (a) the first solvent and
5 the ginseng are combined in a proportion of about 8 ml of first solvent per gram of ginseng.
12. The process of claim 1, wherein in step (c) the water and the first ginseng residue are combined in a proportion of about 7-9 ml of water per gram of ginseng residue.
- 10 13. The process of claim 1, wherein in step (c) the water and the first ginseng residue are combined in a proportion of about 8 ml of water per gram of ginseng residue.
14. Ginseng fraction PQ_2 , produced according to the process of any of claims 1-13.
- 15 15. A process of preparing ginseng fraction PQ_{223} , the process comprising:
- (a) providing ginseng fraction PQ_2 , produced according to the process of any of claims 1-13;
 - (b) fractionating the ginseng fraction PQ_2 to produce a first
20 elution fraction and a second elution fraction, wherein the first elution fraction corresponds to a carbohydrate peak observed between 35 and 50 ml of elution volume and the second elution fraction corresponds to a carbohydrate peak observed between 50 and 85 ml of elution volume,

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as determined by gel filtration chromatography using the following materials:

- (1) a chromatographic column containing a matrix of a spherical cross-linked co-polymer of allyl dextran and N,N'-methylenebisacrylamide, having a bed dimension of 16 × 600 mm, a bed volume of 120 ml, and a fractionation range (MW) of 5000 to 250,000 for globular proteins and 1000 to 80,000 for dextrans, and
- (2) an elution buffer of Tris-HCl containing 0.1 N HCl and 0.3 M NaCl at a pH of 7.0; and
- (c) isolating and combining the first elution fraction and the second elution fraction to produce ginseng fraction PQ₂₂₃.

16. The method of claim 15, wherein ginseng fraction PQ₂ is fractionated using gel filtration chromatography.

17. Ginseng fraction PQ₂₂₃, produced according to the process of claims 15 or 16.

18. A process of preparing ginseng fraction CVT-E002, the process comprising:

- (a) combining American ginseng with a first solvent comprising an alcohol in a proportion of about 7-9 ml of first solvent per gram of ginseng and heating the resulting solution at a temperature of about 80-100°C for a time period of about 2-4 hours, to produce a first ginseng solution;
- (b) thereafter separating the first ginseng solution to produce an alcohol/ginseng solution and a first ginseng residue;
- (c) thereafter combining the first ginseng residue with water in

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a proportion of about 7-9 ml of water per gram of ginseng residue and heating the ginseng residue solution at a temperature of about 80-100°C for a time period of about 2-4 hours, to produce a ginseng residue solution;

5 (d) thereafter separating the ginseng residue solution to produce a second ginseng residue and an aqueous extract solution containing a ginseng extract; and

 (e) drying or concentrating the aqueous extract solution to produce ginseng fraction CVT-E002.

10 19. The process of claim 18, wherein in step (a) the first solvent and the sample are combined in a proportion of about 8 ml of first solvent per gram of sample.

 20. The process of claim 18, wherein in step (c) the water and the first ginseng residue are combined in a proportion of about 8 ml of water per
15 gram of ginseng residue.

 21. The process of claim 18, wherein in step (a) the first ginseng solution is heated for a time period of about 3 hours.

 22. The process of claim 18, wherein in step (c) the ginseng residue solution is heated for a time period of about 3 hours.

20 23. The process of claim 18, wherein the alcohol in the first solvent comprises a saturated or unsaturated C₁-C₆ alcohol.

 24. The process of claim 18, wherein the alcohol in the first solvent

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comprises ethanol or methanol.

25. Ginseng fraction CVT-E002, produced according to the process of any of claims 18-24.

26. A ginseng fraction, having a carbohydrate content which
5 comprises about 2-6 mol% rhamnose, about 41-49 mol% galacturonic acid, about 12-18 mol% glucose, about 16-22 mol% galactose and about 12-19 mol% arabinose.

27. The ginseng fraction of claim 26, wherein the carbohydrate
10 content comprises about 3-5 mol% rhamnose, about 43-47 mol% galacturonic acid, about 14-16 mol% glucose, about 18-20 mol% galactose and about 14-17 mol% arabinose.

28. The ginseng fraction of claim 26, wherein the carbohydrate
15 content comprises about 4 mol% rhamnose, about 45 mol% galacturonic acid, about 15 mol% glucose, about 19 mol% galactose and about 15 mol% arabinose.

29. A ginseng fraction, having a carbohydrate content which
comprises about 3-8 mol% rhamnose, about 36-44 mol% galacturonic acid, about 2-7 mol% glucose, about 25-33 mol% galactose and about 17-25 mol% arabinose.

20 30. The ginseng fraction of claim 29, wherein the carbohydrate content comprises about 4-7 mol% rhamnose, about 37-42 mol% galacturonic acid, about 3-6 mol% glucose, about 27-32 mol% galactose

and about 19-24 mol% arabinose.

31. The ginseng fraction of claim 29, wherein the carbohydrate content comprises about 5 mol% rhamnose, about 39 mol% galacturonic acid, about 4 mol% glucose, about 29 mol% galactose and about 21
5 mol% arabinose.

32. A ginseng fraction, having a carbohydrate content which comprises about 0.5-5 mol% rhamnose, about 11-22 mol% galacturonic acid, about 40-60 mol% glucose, about 10-19 mol% galactose and about 11-19 mol% arabinose.

10 33. The ginseng fraction of claim 32, wherein the carbohydrate content comprises about 1-3 mol% rhamnose, about 13-20 mol% galacturonic acid, about 42-57 mol% glucose, about 12-17 mol% galactose and about 13-17 mol% arabinose.

15 34. A pharmaceutical composition, comprising the ginseng fraction according to any one of claims 14, 17 and 25-33 in combination with a pharmaceutically acceptable carrier.

35. Use of a ginseng fraction according to any one of claims 14, 17 and 25-33, alone or in combination with another medicament, in the preparation of a pharmaceutical composition suitable for treating a
20 condition characterized by low immunity.

36. Use of claim 35, wherein the condition is selected from the group consisting of common cold, influenza, chronic fatigue syndrome, AIDS

and cancer.

37. Use of a ginseng fraction according to any one of claims 14, 17 and 25-33 to stimulate the production of IL-1, IL-6 and/or TNF- α in cells.

38. Use of a ginseng fraction according to any one of claims 14, 17
5 and 25-33 to stimulate the *in vitro* or *in vivo* production of immunoglobulins.

39. Use of a ginseng fraction according to any one of claims 14, 17 and 25-33 to activate B-lymphocyte proliferation and antibody production therefrom.

10 40. A method of treating a condition characterized by low immunity in a patient in need thereof, comprising administering to the patient a condition treating effective amount of a ginseng fraction according to any one of claims 14, 17 and 25-33.

15 41. The method of claim 40, wherein the condition is selected from the group consisting of common cold, influenza, chronic fatigue syndrome, AIDS and cancer.

42. A process of preparing ginseng fraction PQ₂A, the process comprising:

20 (a) providing ginseng fraction PQ₂, produced according to the process of any of claims 1-13;

(b) fractionating the ginseng fraction PQ₂ to produce an elution fraction which corresponds to a carbohydrate peak observed between

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35 and 50 ml of elution volume, as determined by gel filtration chromatography using the following materials:

- (1) a chromatographic column containing a matrix of a spherical cross-linked co-polymer of allyl dextran and N,N'-methylenebisacrylamide, having a bed dimension of 16 × 600 mm, a bed
5 volume of 120 ml, and a fractionation range (MW) of 5000 to 250,000 for globular proteins and 1000 to 80,000 for dextrans, and
- (2) an elution buffer of Tris-HCl containing 0.1 N HCl and 0.3 M NaCl at a pH of 7.0; and
- 10 (c) isolating the elution fraction to produce ginseng fraction PQ₂A.

43. Ginseng fraction PQ₂A, produced according to the process of claim 42.

44. A process of preparing ginseng fraction PQ₂B, the process
15 comprising:

- (a) providing ginseng fraction PQ₂, produced according to the process of any of claims 1-13;
- (b) fractionating the ginseng fraction PQ₂B to produce an elution fraction which corresponds to a carbohydrate peak observed
20 between 50 and 85 ml of elution volume, as determined by gel filtration chromatography using the following materials:

- (1) a chromatographic column containing a matrix of a spherical cross-linked co-polymer of allyl dextran and N,N'-methylenebisacrylamide, having a bed dimension of 16 × 600 mm, a bed
25 volume of 120 ml, and a fractionation range (MW) of 5000 to 250,000 for globular proteins and 1000 to 80,000 for dextrans, and

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(2) an elution buffer of Tris-HCl containing 0.1 N HCl and 0.3 M NaCl at a pH of 7.0; and

(c) isolating the elution fraction to produce ginseng fraction PQ₂B.

5 45. Ginseng fraction PQ₂B, produced according to the process of claim 44.

46. A process of preparing ginseng fraction PQ₂C, the process comprising:

10 (a) providing ginseng fraction PQ₂, produced according to the process of any of claims 1-13;

(b) fractionating the ginseng fraction PQ₂ to produce an elution fraction which corresponds to a carbohydrate peak observed between 95 and 110 ml of elution volume, as determined by gel filtration chromatography using the following materials:

15 (1) a chromatographic column containing a matrix of a spherical cross-linked co-polymer of allyl dextran and N,N'-methylenebisacrylamide, having a bed dimension of 16 × 600 mm, a bed volume of 120 ml, and a fractionation range (MW) of 5000 to 250,000 for globular proteins and 1000 to 80,000 for dextrans, and

20 (2) an elution buffer of Tris-HCl containing 0.1 N HCl and 0.3 M NaCl at a pH of 7.0; and

(c) isolating the elution fraction to produce ginseng fraction PQ₂C.

25 47. Ginseng fraction PQ₂C, produced according to the process of claim 46.

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48. A process of preparing ginseng fraction PQ₂D, the process comprising:

(a) providing ginseng fraction PQ₂, produced according to the process of any of claims 1-13;

5 (b) fractionating the ginseng fraction PQ₂ to produce an elution fraction which corresponds to a carbohydrate peak observed between 120 and 250 ml of elution volume, as determined by gel filtration chromatography using the following materials:

(1) a chromatographic column containing a matrix of a
10 spherical cross-linked co-polymer of allyl dextran and N,N'-methylenebisacrylamide, having a bed dimension of 16 × 600 mm, a bed volume of 120 ml, and a fractionation range (MW) of 5000 to 250,000 for globular proteins and 1000 to 80,000 for dextrans, and

(2) an elution buffer of Tris-HCl containing 0.1 N HCl and
15 0.3 M NaCl at a pH of 7.0; and

(c) isolating the elution fraction to produce ginseng fraction PQ₂D.

49. Ginseng fraction PQ₂D, produced according to the process of claim 48.

20 50. A composition, comprising at least two of the following ginseng fractions:

(a) ginseng fraction PQ₂A, according to claim 43;

(b) ginseng fraction PQ₂B, according to claim 45;

(c) ginseng fraction PQ₂C, according to claim 47; and

25 (d) ginseng fraction PQ₂D, according to claim 49.

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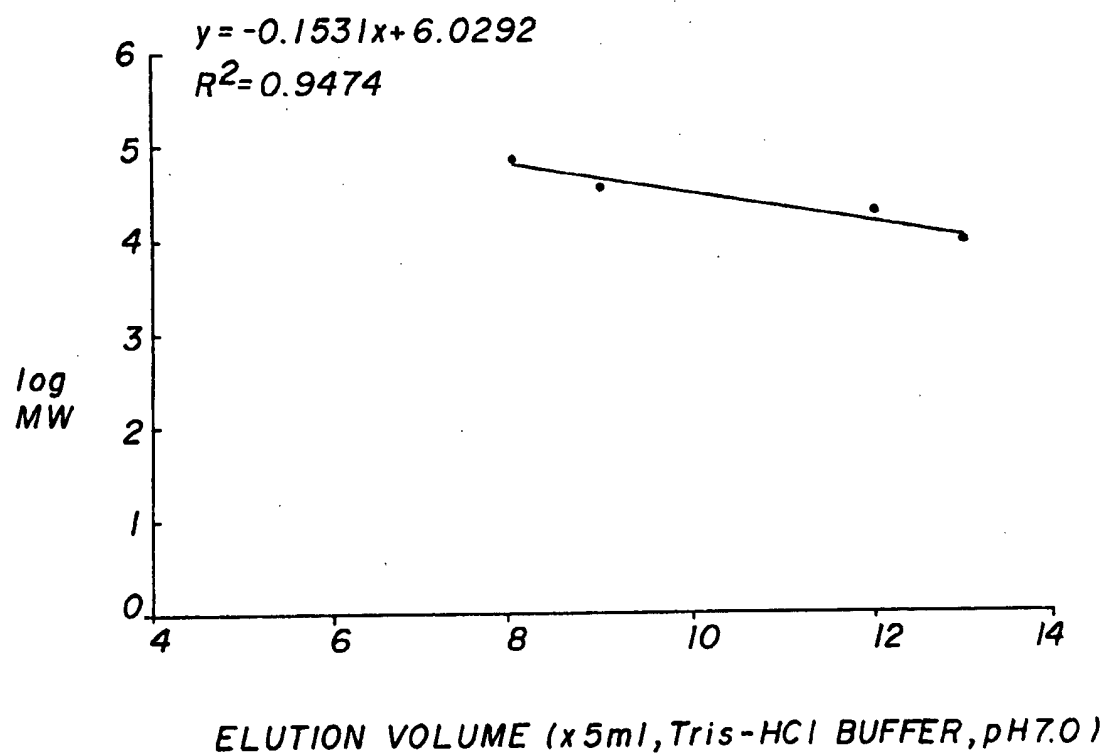


FIG. 1

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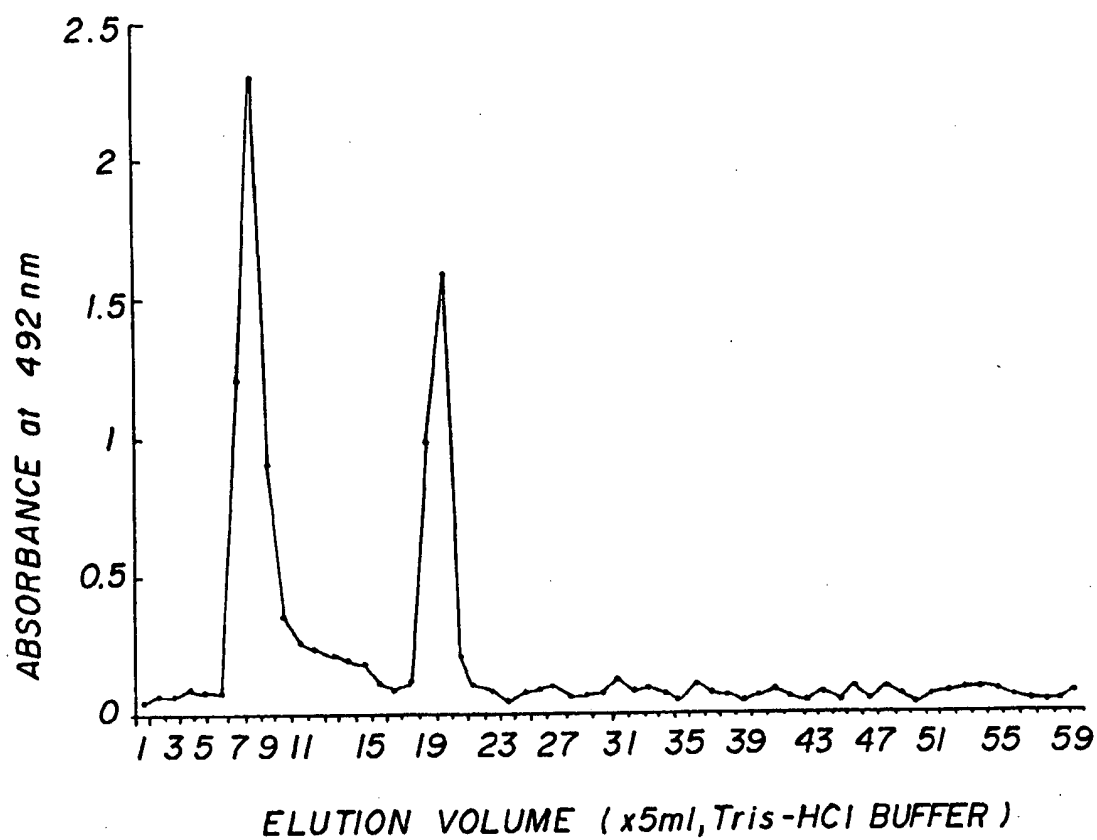


FIG. 2

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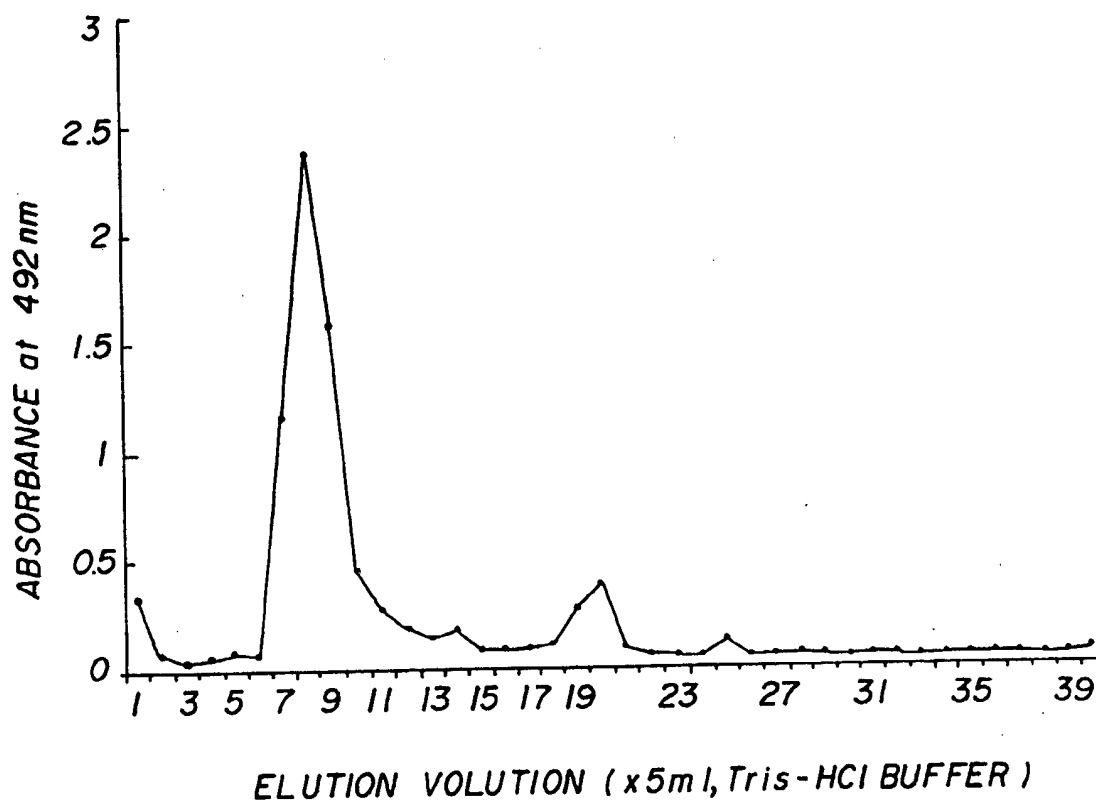


FIG. 3

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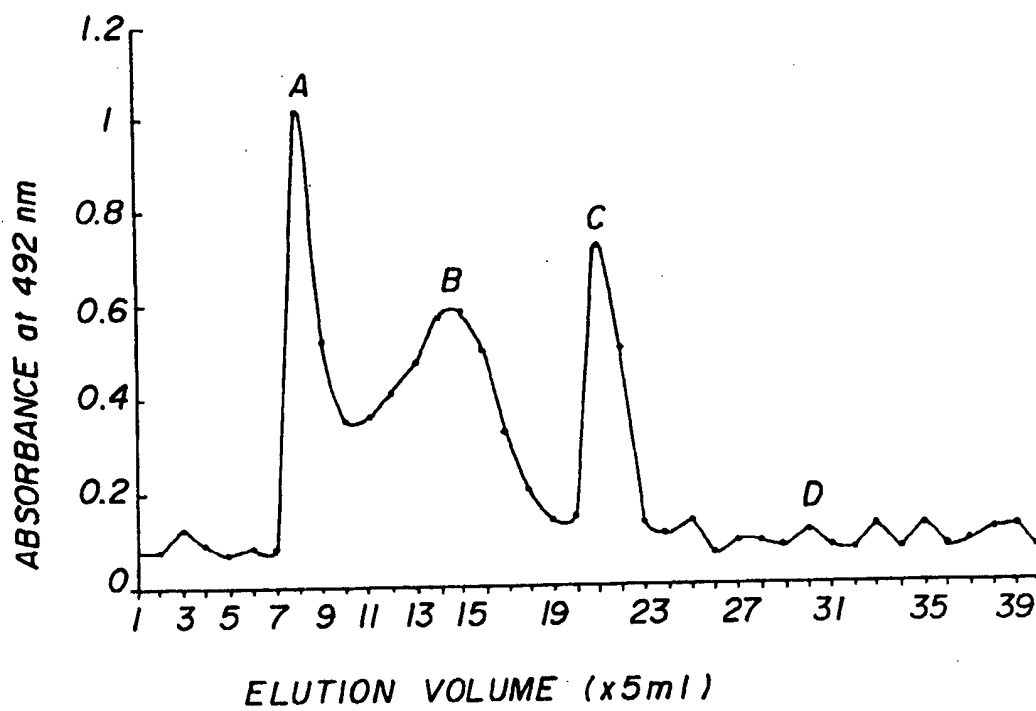


FIG. 4

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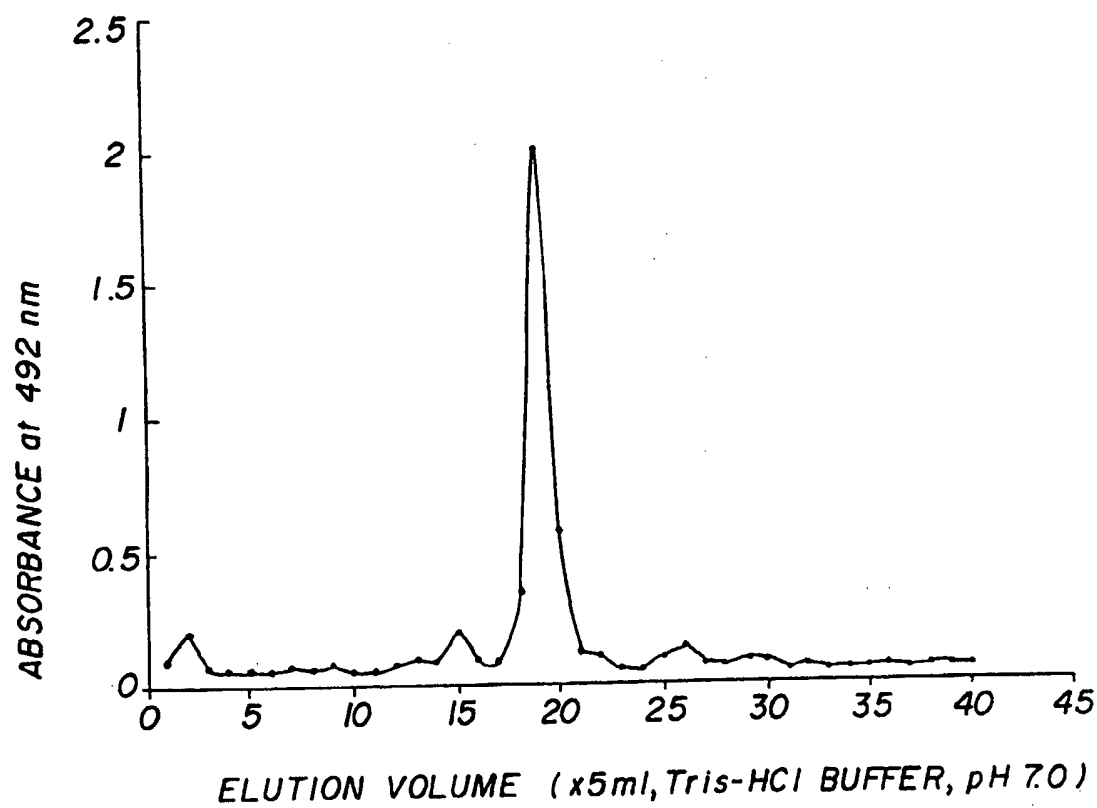


FIG. 5

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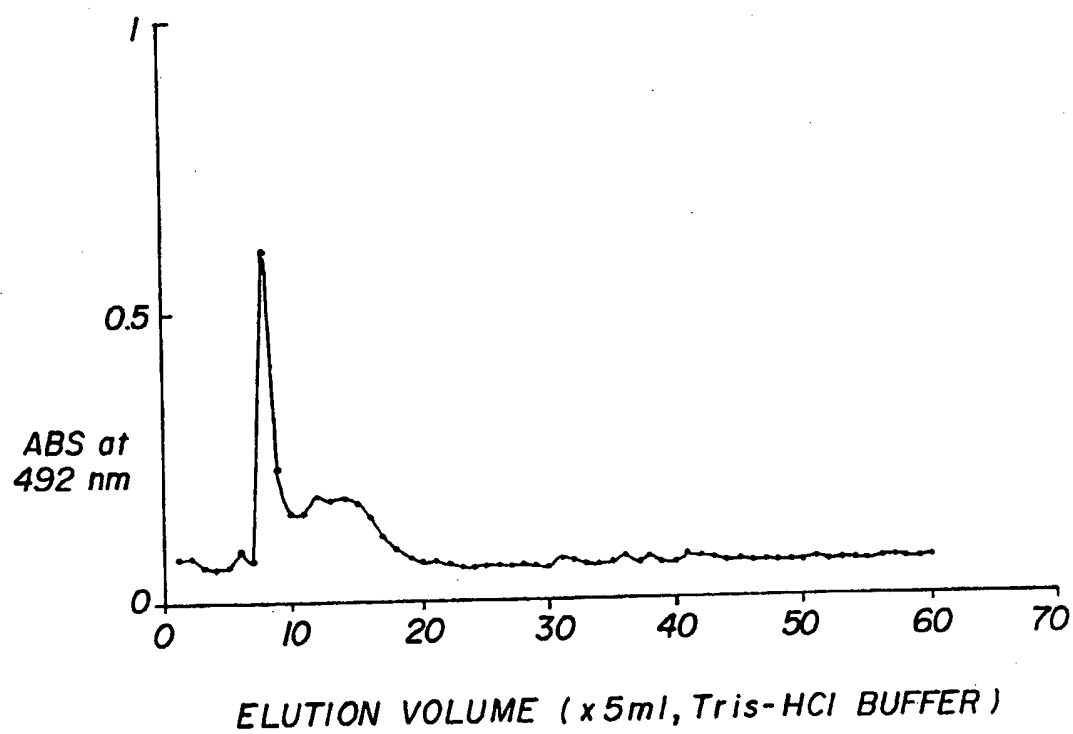


FIG. 6

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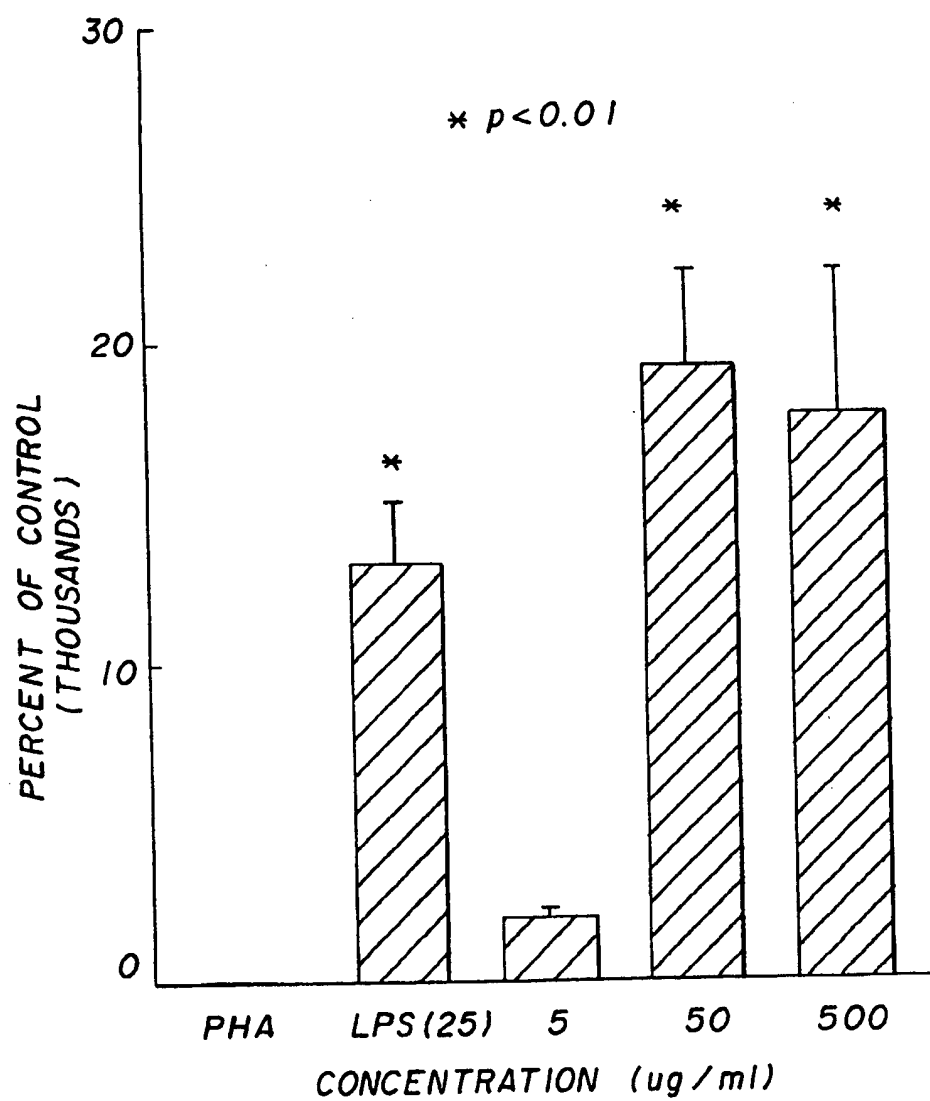


FIG. 7

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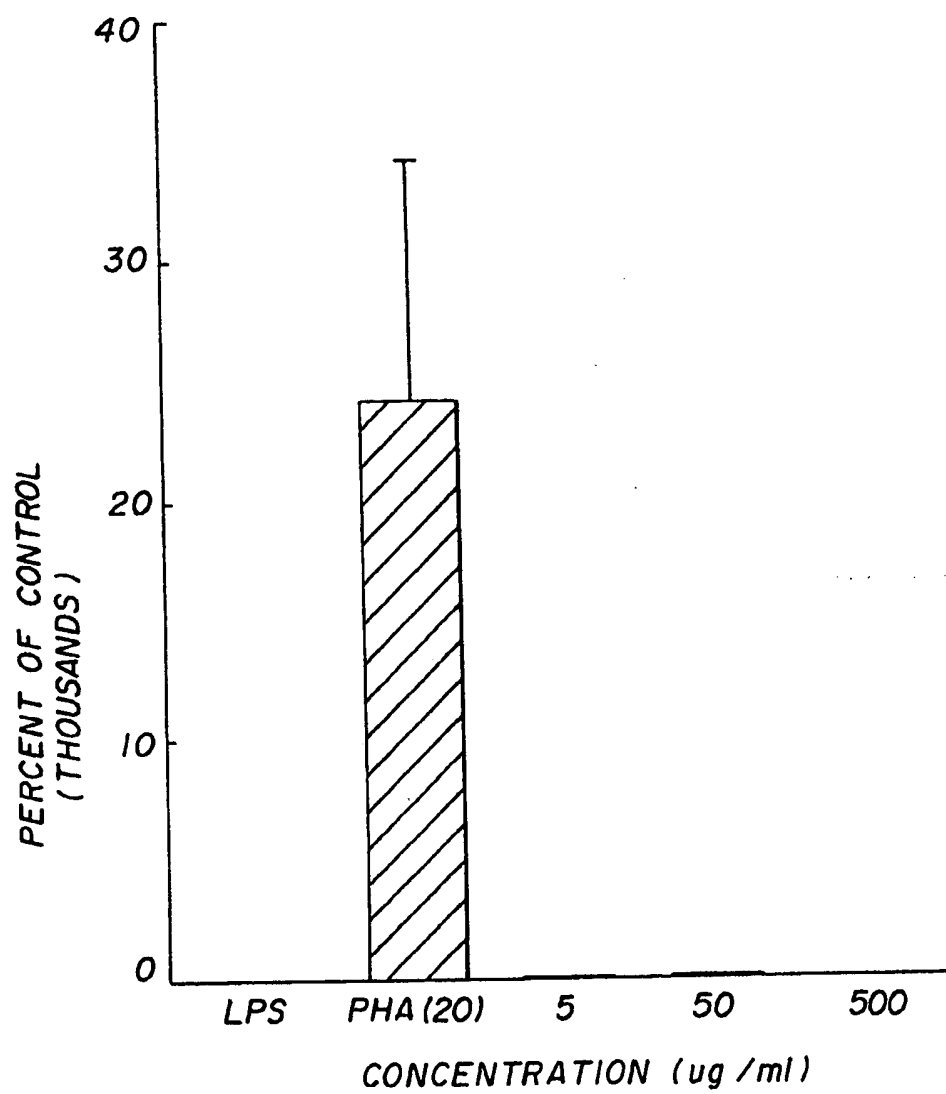


FIG. 8

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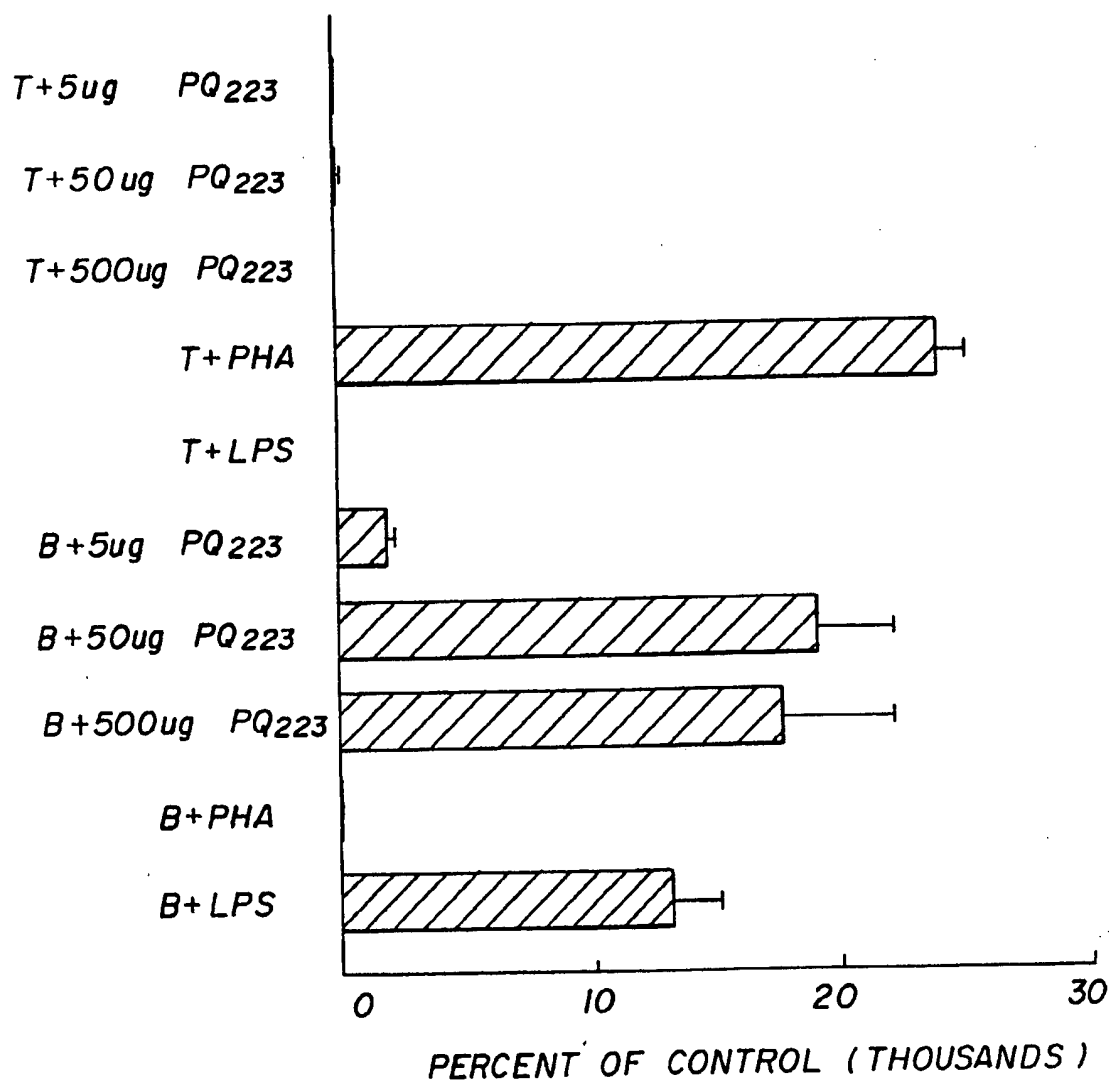


FIG. 9

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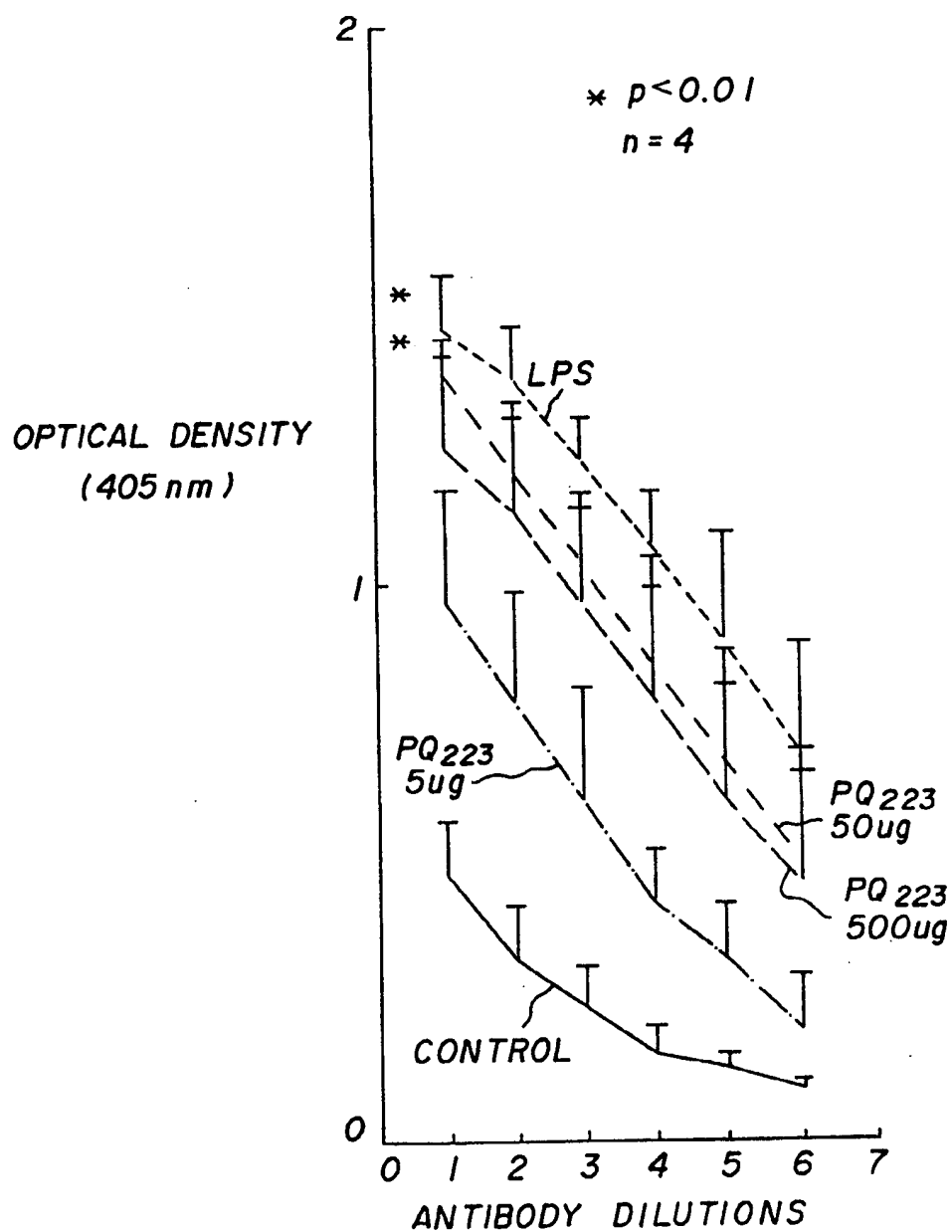


FIG. 10

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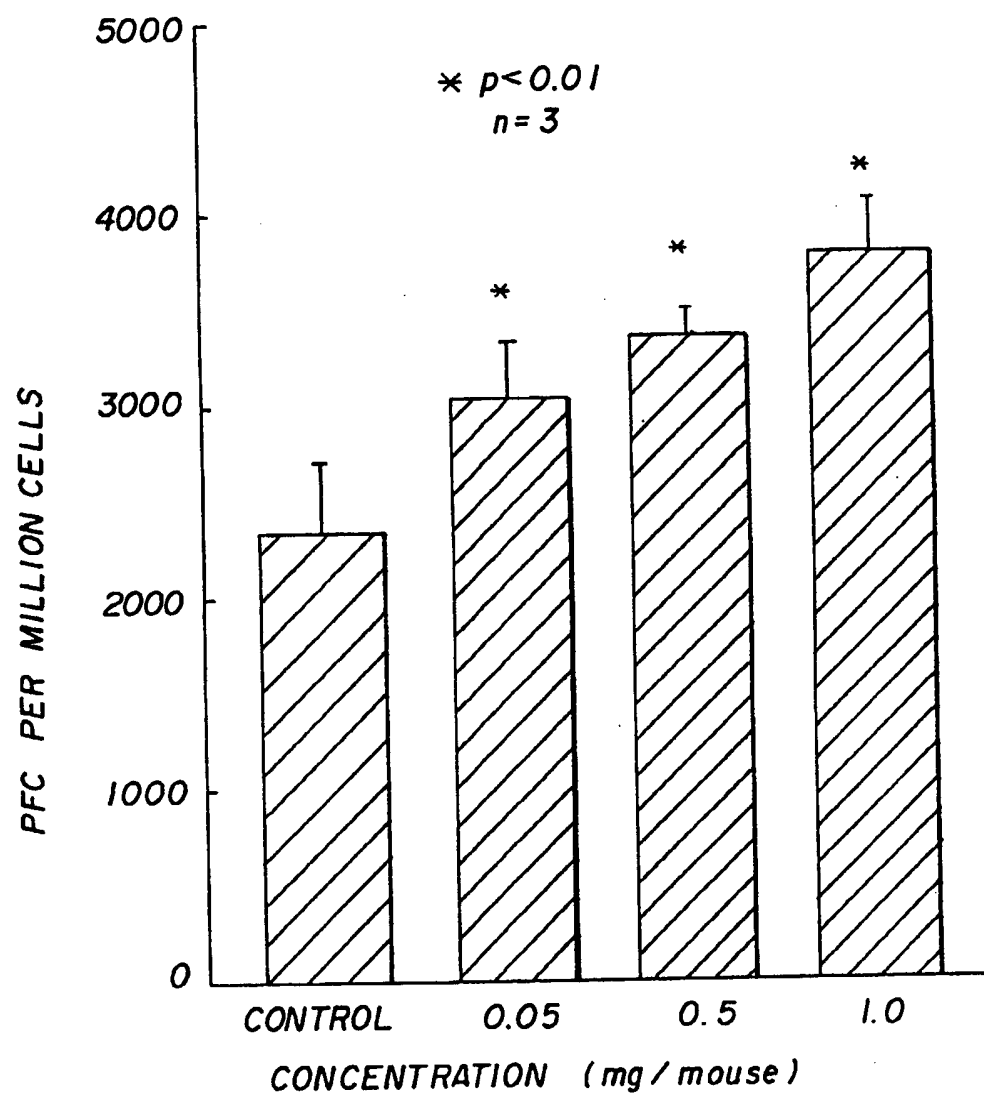


FIG. 11

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US98/25724

A. CLASSIFICATION OF SUBJECT MATTER

IPC(6) :A61K 35/78

US CL :424/195.1

According to International Patent Classification (IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

U.S. : 424/195.1

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practicable, search terms used)

APS, STN databases

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category*	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
X	FUJIMOTO et al. Cytotoxic Acetylenes from Panax Quinquefolium. Chem. Pharm. Bull. February 1991, Vol 39, No. 2, pages 521-523, see entire document.	14

☐ Further documents are listed in the continuation of Box C. ☐ See patent family annex.

* Special categories of cited documents:	*T* later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention
A document defining the general state of the art which is not considered to be of particular relevance	*X* document of particular relevance; the claimed invention cannot be considered novel or cannot be considered to involve an inventive step when the document is taken alone
E earlier document published on or after the international filing date	*Y* document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art
L document which may throw doubts on priority claim(s) or which is cited to establish the publication date of another citation or other special reason (as specified)	*G* document member of the same patent family
O document referring to an oral disclosure, use, exhibition or other means	
P document published prior to the international filing date but later than the priority date claimed	

Date of the actual completion of the international search 05 MARCH 1999	Date of mailing of the international search report 05 APR 1999
Name and mailing address of the ISA/US Commissioner of Patents and Trademarks Box PCT Washington, D.C. 20231 Facsimile No. (703) 305-3230	Authorized officer <i>D. Kussner</i> CHRISTOPHER TATE Telephone No. (703) 308-0196

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US98/25724

Box I Observations where certain claims were found unsearchable (Continuation of item 1 of first sheet)

This international report has not been established in respect of certain claims under Article 17(2)(a) for the following reasons:

1. ☐ Claims Nos.:
because they relate to subject matter not required to be searched by this Authority, namely:
2. ☐ Claims Nos.:
because they relate to parts of the international application that do not comply with the prescribed requirements to such an extent that no meaningful international search can be carried out, specifically:
3. ☒ Claims Nos.: 17 and 34-41
because they are dependent claims and are not drafted in accordance with the second and third sentences of Rule 6.4(a).

Box II Observations where unity of invention is lacking (Continuation of item 2 of first sheet)

This International Searching Authority found multiple inventions in this international application, as follows:

Please See Extra Sheet.

1. ☐ As all required additional search fees were timely paid by the applicant, this international search report covers all searchable claims.
2. ☐ As all searchable claims could be searched without effort justifying an additional fee, this Authority did not invite payment of any additional fee.
3. ☐ As only some of the required additional search fees were timely paid by the applicant, this international search report covers only those claims for which fees were paid, specifically claims Nos.:
4. ☒ No required additional search fees were timely paid by the applicant. Consequently, this international search report is restricted to the invention first mentioned in the claims; it is covered by claims Nos.:

1-14

Remark on Protest

- ☐ The additional search fees were accompanied by the applicant's protest.
☐ No protest accompanied the payment of additional search fees.

INTERNATIONAL SEARCH REPORT

International application No.
PCT/US98/25724

BOX II. OBSERVATIONS WHERE UNITY OF INVENTION WAS LACKING

This ISA found multiple inventions as follows:

This application contains the following inventions or groups of inventions which are not so linked as to form a single inventive concept under PCT Rule 13.1. In order for all inventions to be searched, the appropriate additional search fees must be paid.

- I. Claims 1-14, drawn to a first ginseng extract product, a first method of making such extract product, and a method of using the extract product.
- II. Claims 15-17, drawn to a second ginseng extract product and a second method of making such extract product.
- III. Claims 18-25, drawn to a third ginseng extract product and a third method of making such extract product.
- IV. Claims 26-28, drawn to a fourth ginseng extract product.
- V. Claims 29-31, drawn to a fifth ginseng extract product.
- VI. Claims 32-33, drawn to a sixth ginseng extract product.
- VII. Claims 42-43, drawn to a seventh ginseng extract product and a fourth method of making such extract product.
- IIIX. Claims 44-45, drawn to an eighth ginseng extract product and a fifth method of making such extract product.
- IX. Claims 46-47, drawn to a ninth ginseng extract product and a sixth method of making such extract product.
- X. Claims 48-49, drawn to a tenth ginseng extract product and a seventh method of making such extract product.
- XI. Claim 50, drawn to an eleventh ginseng extract product.

The inventions listed as Groups I-XI do not relate to a single inventive concept under PCT Rule 13.1 because, under PCT Rule 13.2, they lack the same or corresponding special technical features for the following reasons: The products of Groups I-XI are chemically distinct in their structures as are their methods of production, and therefore are not linked so as to form a single inventive concept.